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(21) International Application Number: PCT/CA98/01129 (22) International Filing Date: 11 December 1998 (11.12.98) (30) Priority Data: 60/069,531 12 December 1997 (12.12.97) US (71) Applicant (for all designated States except US): THE UNIVERSITY OF WESTERN ONTARIO [CA/CA]; Stevenson - Lawson Building, Room 319, London, Ontario N6A 5B8 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): SINGH, Bhagirath [CA/CA]; 146 St. Bees Close, London, Ontario N6G 4B8 (CA). RIDER, Beverley [CA/US]; 531 Lake Boulevard, Redwood City, CA 94062 (US). (74) Agent: BERESKIN & PARR; 40th floor, 40 King Street West, Toronto, Ontario M5H 3Y2 (CA).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: NOVEL PEPTIDE, apoEp1.B, COMPOSITIONS AND USES THEREOF (57) Abstract <p>A peptide derived from apolipoprotein E termed apoEp1.B which includes amino acids 239-252 of the apolipoprotein E is described. The apoEp1.B peptide is a potent immune modulator that acts on a variety of immune cells. Interestingly, apoEp1.B is a dual modulator, capable of both inducing and suppressing an immune response. In particular, apoEp1.B has been shown to induce differentiation of stem cells into dendritic cells, to induce tumor cell differentiation and activation, to inhibit inflammation and to inhibit autoimmune disease.</p>		

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Title: NOVEL PEPTIDE, apoEp1.B, COMPOSITIONS AND USES THEREOF

FIELD OF THE INVENTION

The invention relates to methods and compositions for immune modulation.

BACKGROUND OF THE INVENTION

5 The immune system is a complex, multifactorial defense system that protects the body from a wide range of infectious diseases including viruses, bacteria, parasites and fungi. Although critical for our survival, in certain instances, such as autoimmune disease, transplant rejection, allergies and inflammation, the immune system can be the cause of illness. In such instances it is desirable to suppress or tolerize the
10 immune response.

 The immune system is comprised of a large variety of cells derived from undifferentiated hematopoietic stem cells and includes phagocytes (such as neutrophil polymorphs, monocytes, macrophages and dendritic cells) and lymphocytes such as T cells and B cells and natural killer cells.

15 Dendritic cells are interesting immune cells as, depending on the circumstances, they can either activate or suppress an immune response. With regard to immune activation, dendritic cells (DCs) are potent lymphocyte stimulators and are extremely effective antigen presenting cells. Recently, considerable interest has been generated in the potential use of dendritic cells for the therapy of cancer and infectious
20 diseases. DCs pulsed with tumour peptides elicit protective and antitumour immunity in mice (Mayordama et al., 1995). Patients with B cell lymphomas have been successfully vaccinated with autologous antigen-pulsed DC directly isolated from the blood (Hsu et al., 1996). Flt3 ligand, which induces DC maturation, resulted in tumour regression and antitumour immune response in mice (Lynch et al., 1997). Unfortunately, advances in
25 treatment of tumors with DCs have been limited by their trace level *in vivo*. Efforts in this area are directed at increasing DC numbers and level of activation. With regard to tolerance, dendritic cells have recently been shown to be involved in the induction of central as well as peripheral tolerance and may be useful in treating autoimmunity, allergies and transplantation rejection.

30 **SUMMARY OF THE INVENTION**

 The present inventors have prepared a peptide derived from apolipoprotein E termed apoEp1.B which includes amino acids 239-252 of the apolipoprotein E. The inventors have found that surprisingly, apoEp1.B is a potent immune modulator that acts on a variety of immune cells. Interestingly, apoEp1.B is a dual
35 modulator, capable of both inducing and suppressing an immune response. In particular, apoEp1.B has been shown to induce differentiation of immature cells into dendritic cells, to induce tumor cell differentiation and activation, to inhibit inflammation and to inhibit autoimmune disease.

Accordingly, the present invention provides an isolated apoEp1.B peptide comprising amino acids 239-252 of an apolipoprotein E protein. In a preferred embodiment, the present invention provides an isolated apoEp1.B peptide having the amino acid sequence TQQIRLQAEIFQAR (murine) or AQQIRLQAEAFQAR (human). The invention
5 also includes analogs, fragments, elongations and derivatives of a peptide of the invention. Analogs and derivatives of the peptides include peptides having the following sequences: TAQIRLQAEIFQAR; TQAIRLQAEIFQAR; TQQARLQAEIFQAR and TQQIALQAEIFQAR. Fragments and elongations of the peptides include peptides that have the following sequences: QTQQIRLQAEIFQAR and QQIRLQAEIFQAR. The present
10 invention also provides a nucleic acid molecule encoding the apoEp1.B peptide, or an analog, fragment or derivative thereof.

The present invention further provides a method of immune modulation comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or animal in need thereof.

15 According to one embodiment, the peptide can induce immune tolerance. In particular, the present inventors have demonstrated that the apoEp1.B peptide can activate monocytes to differentiate into tolerogenic dendritic cells. The induction of tolerogenic dendritic cells can have a wide variety of therapeutic applications including inflammation, autoimmune disease and transplantation.

20 In another embodiment, the apoEp1.B peptide is useful in inhibiting inflammation. In a preferred embodiment, the peptide can inhibit atherosclerotic plaque formation *in vivo*.

In a further embodiment, the apoEp1.B peptide can be used to prevent or treat an autoimmune reaction or disease. The present inventors have also demonstrated
25 that the apoEp1.B peptide can protect NOD mice from developing diabetes. In a preferred embodiment, the autoimmune disease is diabetes.

In a further aspect, the apoEp1.B peptide can be used to induce an immune response by activating immune cells. In one embodiment, the peptide, in combination with other cytokines such as IL-4, GM-CSF, TNF α and Flt3 ligand may induce immature
30 dendritic cells to differentiate into mature immunogenic dendritic cells. Mature dendritic cells can be used in a wide variety of applications including tumor immunotherapy.

In another aspect, the apoEp1.B peptide can be used to treat tumors of immune origin by inducing their differentiation. In particular, the present inventors have demonstrated that the apoEp1.B peptide can induce the differentiation and activation of
35 monocytic, monoblastic leukemia and lymphoma tumor cells.

Other features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples while indicating preferred

embodiments of the invention are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

5 The invention will now be described in relation to the drawings in which:
 Figure 1A and B are photographs of spleen cells incubated with apoEp1.B (B) or a control peptide (A) for 48 hours.

 Figure 1C and D are photographs of PU5-1.8 cells incubated with apoEp1.B (D) or a control peptide (C) for 48 hours.

10 Figure 2(A-D) is a FACS analysis of J77A cells incubated with apoEp1.B (B and D) or with a control peptide (A and C) for 48 hours.

 Figure 3 is a FACS analysis of PU5-1.8 cells induced with apoEp1.B or apoEp1.D and stained with various markers.

 Figure 4 shows FACS analysis of BALB/c splenocytes incubated with
15 apoEp1.B or apoEp1.D.

 Figure 5 shows a FACS analysis illustrating that apoEp1.B induced DC surface molecule expression is dose-dependent.

 Figure 6 shows a FACS analysis which demonstrates that apoEp1.B is not species-specific.

20 Figure 7 are bar graphs demonstrating chemokine production from PU5-1.8 cells incubated with apoEp1.B, apoEp1 and a control.

 Figure 8A is a FACS analysis showing SCC and FSC profiles of apoEp1.D and apoEp1.B primed PEC cells.

 Figure 8B is a FACS analysis showing surface marker expression of
25 apoEp1.D (unlabelled peak) and apoEp1.B (labelled peak) primed PEC cells.

 Figure 9 is a bar graph showing activation of spleen cells in the presence of apoEp1.B and/or ConA.

 Figure 10 shows a section of iliofemoral arteries: (A) untreated, no surgery; (B) untreated, surgery and (C) treated with apoEp1.B and surgery (arterial section shown).

30 Figure 11 shows histograms which illustrate the induction of Th2-like cells by apoEp1.B immunization.

 Figure 12 shows a histogram which illustrates allostimulatory abilities of apoEp1.B treated cells.

 Figure 13A is a graph which illustrates apoEp1.B protection of NOD mice
35 from spontaneous diabetes.

 Figure 13B is a graph which illustrate apoEp1.D protection of NOD mice from adoptively transferred diabetes.

Figure 14 is a graph showing the percentage of mice protected from diabetes versus time, in the presence and absence of apoEp1.B.

Figure 15A-D illustrate the effect of different concentrations of apoEp1.B on proliferation of U-937 cells.

5 Figure 16 shows a FACS analysis which illustrates that single amino acid deletions or elongations decrease the activity of apoEp1.B.

DETAILED DESCRIPTION OF THE INVENTION

As hereinbefore mentioned, the present inventors have prepared a peptide from both human and murine apolipoprotein E termed apoEp1.B (239-252) which is a potent
10 immune modulator and can be used in a wide variety of applications.

I. PEPTIDES OF THE INVENTION

Broadly stated, the present invention provides an isolated apoEp1.B peptide comprising amino acids 239-252 of an apolipoprotein E protein or an analog, fragment, elongation or derivative thereof.

15 In one aspect, the present invention provides an isolated apoEp1.B peptide having the amino acid sequence TQQIRLQAEIFQAR (murine)(SEQ. ID. No. 1) or AQQIRLQAEAFQAR (human)(SEQ. ID. No. 2) or an analog, fragment, elongation or derivative of the peptide. The invention also includes a nucleic acid molecule encoding the apoEp1.B peptide, or an analog, fragment, elongation or derivative thereof.

20 The term "analog" includes any peptide having an amino acid residue sequence substantially identical to the human or mouse apoEp1.B sequence specifically shown herein in which one or more residues have been conservatively substituted with a functionally similar residue and which displays the ability to mimic apoEp1.B as described herein. Examples of conservative substitutions include the substitution of one non-
25 polar (hydrophobic) residue such as alanine, isoleucine, valine, leucine or methionine for another, the substitution of one polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, between glycine and serine, the substitution of one basic residue such as lysine, arginine or histidine for another, or the substitution of one acidic residue, such as aspartic acid or glutamic acid for another. The
30 phrase "conservative substitution" also includes the use of a chemically derivatized residue in place of a non-derivatized residue provided that such polypeptide displays the requisite activity.

Analogous of the peptides include peptides having the following sequences:
TAQIRLQAEIFQAR (SEQ.ID.NO.:3); TQAIRLQAEIFQAR (SEQ.ID.NO.:4);
35 TQQARLQAEIFQAR (SEQ.ID.NO.:5) and TQQIALQAEIFQAR (SEQ.ID.NO.:6).

"Derivative" refers to a peptide having one or more residues chemically derivatized by reaction of a functional side group. Such derivatized molecules include for example, those molecules in which free amino groups have been derivatized to form amine

hydrochlorides, p-toluene sulfonyl groups, carbobenzoxy groups, t-butyloxycarbonyl groups, chloroacetyl groups or formyl groups. Free carboxyl groups may be derivatized to form salts, methyl and ethyl esters or other types of esters or hydrazides. Free hydroxyl groups may be derivatized to form O-acyl or O-alkyl derivatives. The imidazole nitrogen of histidine may be derivatized to form N-im-benzylhistidine. Also included as derivatives are those peptides which contain one or more naturally occurring amino acid derivatives of the twenty standard amino acids. For examples: 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine; and ornithine may be substituted for lysine. Polypeptides of the present invention also include any polypeptide having one or more additions and/or deletions or residues relative to the sequence of a polypeptide whose sequence is shown herein, so long as the requisite activity is maintained.

The term "fragment" refers to any subject peptide having an amino acid residue sequence shorter than that of a peptide whose amino acid residue sequence is shown herein.

The term "elongation" refers to any subject peptide having an amino acid sequence longer by one or two amino acids (either at the carboxy or amino terminal end) than that of a peptide of the present invention. Preferably, the elongation occurs at the amino terminal end.

Fragments and elongations of the peptides include peptides that have the following sequences: QTQQIRLQAEIFQAR (SEQ.ID.NO.:7) and QQIRLQAEIFQAR (SEQ.ID.NO.:8).

The term "apoEp1.B peptide" or "peptide of the invention" as used herein includes a peptide comprising amino acid residues 239 to 252 of an apolipoprotein E protein and includes all analogs, fragments, elongations or derivatives of the apoEp1.B peptide including the sequences provided above in SEQ.ID.NOS.:1-8. Preferably, the apoEp1.B is the murine apoEp1.B sequence TQQIRLQAEIFQAR (SEQ. ID. No. 1) or the human apoEp1.B sequence AQQIRLQAEAFQAR (SEQ. ID. No. 2), .

The apoEp1.B peptide may be modified to make it more therapeutically effective or suitable. For example, it may be cyclized as cyclization allows a peptide to assume a more favourable conformation. Cyclization of the apoEp1.B peptide may be achieved using techniques known in the art. In particular, disulphide bonds may be formed between two appropriately spaced components having free sulfhydryl groups. The bonds may be formed between side chains of amino acids, non-amino acid components or a combination of the two. In addition, the apoEp1.B peptide of the present invention may be converted into pharmaceutical salts by reacting with inorganic acids including hydrochloric acid, sulphuric acid, hydrobromic acid, phosphoric acid, etc., or organic acids

including formic acid, acetic acid, propionic acid, glycolic acid, lactic acid, pyruvic acid, oxalic acid, succinic acid, malic acid, tartaric acid, citric acid, benzoic acid, salicylic acid, benzenesulphonic acid, and tolunesulphonic acids.

5 The apoEp1.B proteins of the invention may also be prepared by conventional techniques. For example, the peptides may be synthesized by chemical synthesis using techniques well known in the chemistry of proteins such as solid or solution phase synthesis (see for example J.M. Stewart, and J.D. Young, Solid Phase Peptide Synthesis, 2nd Ed., Pierce Chemical Co., Rockford Ill. (1984) and G. Barany and R.B. Merrifield, The Peptides: Analysis Synthesis, Biology editors E. Gross and J. Meienhofer
10 Vol. 2 Academic Press, New York, 1980, pp. 3-254 for solid phase synthesis techniques; and M Bodansky, Principles of Peptide Synthesis, Springer-Verlag, Berlin 1984, and E. Gross and J. Meienhofer, Eds., The Peptides: Analysis, Synthesis, Biology, supra, Vol 1, for classical solution synthesis. and Merrifield, 1964, J. Am. Chem. Assoc. 85:2149-2154) or synthesis in homogenous solution (Houbenweyl, 1987, Methods of Organic Chemistry, ed. E.
15 Wansch, Vol. 15 I and II, Thieme, Stuttgart).

The apoEp1.B peptides of the invention may also be produced by recombinant DNA technology. To prepare the peptides of the invention by recombinant DNA techniques, a DNA sequence encoding the apoEp1.B peptide must be prepared. Consequently, the present invention also provides purified and isolated nucleic acid having
20 a nucleotide sequence encoding an apoEp1.B peptide comprising an amino acid sequence TQQIRLQAEIFQAR (SEQ. ID. No. 1) or an amino acid sequence AQQIRLQAEAFQAR (SEQ. ID. No. 2).

The present invention also provides an expression vector comprising a DNA molecule encoding an apoEp1.B peptide adapted for transfection or transformation of a host
25 cell.

Accordingly, the nucleic acid molecules of the present invention may be incorporated in a known manner into an appropriate expression vector which ensures expression of the protein. Possible expression vectors include but are not limited to cosmids, plasmids, or modified viruses (e.g. replication defective retroviruses, adenoviruses and
30 adeno-associated viruses). The vector should be compatible with the host cell used. The expression vectors are "suitable for transformation of a host cell", which means that the expression vectors contain a nucleic acid molecule of the invention and regulatory sequences selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid molecule. Operatively linked is intended to mean that the nucleic acid
35 is linked to regulatory sequences in a manner which allows expression of the nucleic acid.

The invention therefore contemplates a recombinant expression vector of the invention containing a nucleic acid molecule of the invention, or a fragment thereof, and the

necessary regulatory sequences for the transcription and translation of the inserted protein-sequence.

Suitable regulatory sequences may be derived from a variety of sources, including bacterial, fungal, viral, mammalian, or insect genes (For example, see the regulatory sequences described in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Selection of appropriate regulatory sequences is dependent on the host cell chosen as discussed below, and may be readily accomplished by one of ordinary skill in the art. Examples of such regulatory sequences include: a transcriptional promoter and enhancer or RNA polymerase binding sequence, a ribosomal binding sequence, including a translation initiation signal. Additionally, depending on the host cell chosen and the vector employed, other sequences, such as an origin of replication, additional DNA restriction sites, enhancers, and sequences conferring inducibility of transcription may be incorporated into the expression vector.

The recombinant expression vectors of the invention may also contain a selectable marker gene which facilitates the selection of host cells transformed or transfected with a recombinant molecule of the invention. Examples of selectable marker genes are genes encoding a protein such as G418 and hygromycin which confer resistance to certain drugs, β -galactosidase, chloramphenicol acetyltransferase, firefly luciferase, or an immunoglobulin or portion thereof such as the Fc portion of an immunoglobulin preferably IgG.

Recombinant expression vectors can be introduced into host cells to produce a transformant host cell. The term "transformant host cell" is intended to include prokaryotic and eukaryotic cells which have been transformed or transfected with a recombinant expression vector of the invention. The terms "transformed with", "transfected with", "transformation" and "transfection" are intended to encompass introduction of nucleic acid (e.g. a vector) into a cell by one of many possible techniques known in the art. Prokaryotic cells can be transformed with nucleic acid by, for example, electroporation or calcium-chloride mediated transformation. Nucleic acid can be introduced into mammalian cells via conventional techniques such as calcium phosphate or calcium chloride co-precipitation, DEAE-dextran mediated transfection, lipofectin, electroporation or microinjection. Suitable methods for transforming and transfecting host cells can be found in Sambrook et al. (*Molecular Cloning: A Laboratory Manual*, 2nd Edition, Cold Spring Harbor Laboratory press (1989)), and other laboratory textbooks.

Suitable host cells include a wide variety of prokaryotic and eukaryotic host cells. For example, the proteins of the invention may be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus), yeast cells or mammalian cells. Yeast and fungi host cells suitable for carrying out the present invention include, but are not limited to *Saccharomyces cerevisiae*, the genera *Pichia* or *Kluyveromyces* and various species of the

genus *Aspergillus*. Mammalian cells suitable for carrying out the present invention include, among others: COS (e.g., ATCC No. CRL 1650 or 1651), BHK (e.g. ATCC No. CRL 6281), CHO (ATCC No. CCL 61), HeLa (e.g., ATCC No. CCL 2), 293 (ATCC No. 1573) and NS-1 cells. Other suitable host cells can be found in Goeddel, Gene Expression Technology:

5 Methods in Enzymology 185, Academic Press, San Diego, CA (1991).

Another aspect of the invention provides a nucleotide sequence which hybridizes under high stringency conditions to a nucleic acid sequence which encodes an apoEp1.B peptide of the invention. Appropriate stringency conditions which promote DNA hybridization are known to those skilled in the art, or can be found in Current
10 Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1 6.3.6. For example, 6.0 x sodium chloride/sodium citrate (SSC) at about 45°C, followed by a wash of 2.0 x SSC at 50°C may be employed. The stringency may be selected based on the conditions used in the wash step. By way of example, the salt concentration in the wash step can be selected from a high stringency of about 0.2 x SSC at 50°C. In addition, the temperature in the wash
15 step can be at high stringency conditions, at about 65°C.

II. UTILITY OF THE PEPTIDES

A. Therapeutic Methods

The inventors have surprisingly found that the apoEp1.B peptide of the invention is a potent immune modulator that acts on a variety of immune cells.
20 Interestingly, apoEp1.B is a dual modulator, capable of both inducing and suppressing an immune response. As mentioned previously, the apoEp1.B peptide comprises amino acids 239-252 of full length apolipoprotein E (apoE) including fragments, elongations, analogs and derivatives of the peptide. The inventors have shown that apoEp1.B but not apoE or a peptide comprising amino acids 237-250 of apoE can induce the activation and
25 differentiation of immune cells.

Accordingly, the present invention provides a method of immune modulation comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or animal in need thereof.

The apoEp1.B peptide can be used to induce immune tolerance. In
30 particular, the present inventors have demonstrated that the apoEp1.B peptide can activate immature cells to differentiate into tolerogenic dendritic cells. Accordingly, the present invention provides a method of inducing immune tolerance comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or animal in need thereof. The induction of tolerogenic dendritic
35 cells can have a wide variety of therapeutic applications including inflammation, autoimmune disease and transplantation. The tolerogenic dendritic cells can be induced *in vitro* and then transferred to a recipient requiring the cells. Alternatively, the tolerogenic cells may be directly induced *in vivo*.

The inventors have shown that the apoEp1.B peptide is useful in inhibiting inflammation. Accordingly, the present invention provides a method for inhibiting inflammation comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or an animal in need thereof. In a preferred embodiment, the peptide can inhibit atherosclerotic plaque formation *in vivo*. Other inflammatory diseases that may be treated using the apoEp1.B peptide or nucleic acid encoding the apoEp1.B peptide include, arthritis, inflammatory bowel disease (IBD), Sjogren's syndrome, atherosclerosis, restenosis, transplant rejection, transplant vasculopathy, asthma, acute respiratory distress syndrome, allergy, psoriasis, multiple sclerosis, systemic lupus, acute glomerulonephritis, spinal cord trauma, and others.

In a further embodiment, the apoEp1.B peptide can be used to prevent an autoimmune reaction or disease. The present inventors have demonstrated that the apoEp1.B peptide can protect NOD mice from developing diabetes. Accordingly, the present invention provides a method for treating or preventing an autoimmune disease comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to an animal in need thereof. In a preferred embodiment, the autoimmune disease is diabetes. Other autoimmune diseases which may be treated using the apoEp1.B peptide or nucleic acid encoding the apoEp1.B peptide include multiple sclerosis, EAE which is the mouse model of multiple sclerosis, rheumatoid arthritis, Sjogren's syndrome, lupus (SLE), autoimmune thyroid disease and others.

In a further aspect, the apoEp1.B peptide can be used to induce an immune response by activating immune cells. Accordingly, the present invention provides a method of inducing an immune response comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to an animal in need thereof. In one embodiment, the peptide, in combination with other cytokines such as IL-4, GM-CSF, TNF α and Flt3 ligand induce immature dendritic cells to differentiate into mature immunogenic dendritic cells. Mature dendritic cells can be used in a wide variety of applications including tumor immunotherapy.

In another aspect, the apoEp1.B peptide can be used to treat tumors of immune origin by inducing their differentiation. In particular, the present inventors have demonstrated that the apoEp1.B peptide can induce the differentiation and activation of monocytic, monoblastic leukemia and lymphoma tumor cells. Accordingly, the present invention provides a method of treating a tumor comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to an animal in need thereof.

The apoEp1.B peptide or nucleic acid encoding the apoEp1.B peptide may also be used to treat or prevent other diseases or conditions requiring immune activation (including infectious diseases such as viral infections) and immune tolerance (including

tissue or organ transplantation, allergies and the above noted inflammatory and autoimmune diseases).

Administration of an "effective amount" of the apoEp1.B peptide or nucleic acid of the present invention is defined as an amount effective, at dosages and for periods of time necessary to achieve the desired result. The effective amount of a compound of the invention may vary according to factors such as the disease state, age, sex, and weight of the animal. Dosage regimens may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation. The term "animal" as used herein includes all members of the animal kingdom including humans.

In all of the above described therapeutic methods, the apoEp1.B peptide may be administered *in vivo* or *ex vivo*. In *ex vivo* applications, the apoEp1.B peptide may be administered to cells that have been removed from the patient in an *in vitro* culture. After incubating the cells and peptide for a period of time sufficient for the desired effect, the cells may be re-introduced into the patient's body. In one example, monocytes may be removed from a patient and cultured with apoEp1.B to allow them to mature. In addition, tumor antigens or autoantigens may be added when treating cancer or autoimmune diseases, respectively. The mature monocytes expressing antigen can be re-introduced into the patient and will induce an immune response to the tumor or autoantigen.

B. Pharmaceutical Compositions

The present invention includes pharmaceutical compositions containing the apoEp1.B peptide or nucleic acid or substances which modulate the effects of apoEp1.B for use in the described methods for modulating the immune response.

Such pharmaceutical compositions can be for intralesional, intravenous, topical, rectal, parenteral, local, inhalant or subcutaneous, intradermal, intramuscular, intrathecal, transperitoneal, oral, and intracerebral use. The composition can be in liquid, solid or semisolid form, for example pills, tablets, creams, gelatin capsules, capsules, suppositories, soft gelatin capsules, gels, membranes, tubelets, solutions or suspensions. The apoEp1.B peptide is preferably injected in a saline solution either intravenously, intraperitoneally or subcutaneously.

Several modes of administration are available when using a composition containing a nucleic acid molecule encoding an apoEp1.B protein. Recombinant molecules comprising a nucleic acid sequence encoding an apoEp1.B protein, or fragment thereof, may be directly introduced into cells or tissues *in vivo* using delivery vehicles such as retroviral vectors, adenoviral vectors and DNA virus vectors. They may also be introduced into cells *in vivo* or *in vitro* using physical techniques such as microinjection and electroporation or chemical methods such as coprecipitation and incorporation of DNA into liposomes.

Recombinant molecules may also be delivered in the form of an aerosol or by lavage. The nucleic acid molecules of the invention may also be applied extracellularly such as by direct injection into cells.

The pharmaceutical compositions of the invention can be intended for administration to humans or animals. Dosages to be administered depend on individual needs, on the desired effect and on the chosen route of administration.

The pharmaceutical compositions can be prepared by per se known methods for the preparation of pharmaceutically acceptable compositions which can be administered to patients, and such that an effective quantity of the active substance is combined in a mixture with a pharmaceutically acceptable vehicle. Suitable vehicles are described, for example, in Remington's Pharmaceutical Sciences (Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pa., USA 1985).

On this basis, the pharmaceutical compositions include, albeit not exclusively, the active compound or substance in association with one or more pharmaceutically acceptable vehicles or diluents, and contained in buffered solutions with a suitable pH and iso-osmotic with the physiological fluids. The pharmaceutical compositions may additionally contain other agents to enhance the efficacy of the apoEp1.B peptide or nucleic acid.

C. Peptide Mimetics

The present invention also include peptide mimetics of the apoEp1.B peptides of the invention. For example, a peptide derived from a binding domain of apoEp1.B will interact directly or indirectly with an associated molecule in such a way as to mimic the native binding domain. Such peptides may include competitive inhibitors, enhancers, peptide mimetics, and the like. All of these peptides as well as molecules substantially homologous, complementary or otherwise functionally or structurally equivalent to these peptides may be used for purposes of the present invention.

"Peptide mimetics" are structures which serve as substitutes for peptides in interactions between molecules (See Morgan et al (1989), Ann. Reports Med. Chem. 24:243-252 for a review). Peptide mimetics include synthetic structures which may or may not contain amino acids and/or peptide bonds but retain the structural and functional features of a peptide, or enhancer or inhibitor of the invention. Peptide mimetics also include peptoids, oligopeptoids (Simon et al (1972) Proc. Natl. Acad. Sci USA 89:9367); and peptide libraries containing peptides of a designed length representing all possible sequences of amino acids corresponding to a peptide of the invention.

Peptide mimetics may be designed based on information obtained by systematic replacement of L-amino acids by D-amino acids, replacement of side chains with groups having different electronic properties, and by systematic replacement of peptide bonds with amide bond replacements. Local conformational constraints can also be introduced to

determine conformational requirements for activity of a candidate peptide mimetic. The mimetics may include isosteric amide bonds, or D-amino acids to stabilize or promote reverse turn conformations and to help stabilize the molecule. Cyclic amino acid analogues may be used to constrain amino acid residues to particular conformational states. The mimetics can also include mimics of inhibitor peptide secondary structures. These structures can model the 3-dimensional orientation of amino acid residues into the known secondary conformations of proteins. Peptoids may also be used which are oligomers of N-substituted amino acids and can be used as motifs for the generation of chemically diverse libraries of novel molecules.

Peptides of the invention may also be used to identify lead compounds for drug development. The structure of the peptides described herein can be readily determined by a number of methods such as NMR and X-ray crystallography. A comparison of the structures of peptides similar in sequence, but differing in the biological activities they elicit in target molecules can provide information about the structure-activity relationship of the target. Information obtained from the examination of structure-activity relationships can be used to design either modified peptides, or other small molecules or lead compounds which can be tested for predicted properties as related to the target molecule. The activity of the lead compounds can be evaluated using assays similar to those described herein.

Information about structure-activity relationships may also be obtained from co-crystallization studies. In these studies, a peptide with a desired activity is crystallized in association with a target molecule, and the X-ray structure of the complex is determined. The structure can then be compared to the structure of the target molecule in its native state, and information from such a comparison may be used to design compounds expected to possess desired activities.

The following non-limiting examples are illustrative of the present invention:

EXAMPLES

Example 1

30 ApoEp1.B Induces Dendritic Cells and Differentiation of Tumor Cells

Materials and Methods

Reagents

Phorbol, 12-myristate, 13-acetate (PMA) (Calbiochem, La Jolla, CA) was dissolved in ethanol and stored at -80°C.

35 Mice

C57BL/6J(B6) (H2b), apoE knockout (H2b), and BALB/c (H2d) mice between 8 to 18 weeks used in this study were purchased from Jackson Laboratories, Bar

Harbor, ME. Mice were fed a regular mouse chow (#5012), low in fat (4.5% wt/wt) and cholesterol (0.022% wt/wt) (Ralston, Purina, St. Louis, MO.).

Cells and Culture Medium

Murine monocytic PU5-1.8 and J77A1.4 transformed cell lines, human
5 monoblastic leukemia THP-1 and U-937, and murine B cell lymphoma A20 cells were cultured in RPMI 1640 medium (Gibco Laboratories, Grand Island, NY) containing 5×10^5 M 2ME, 10 mM HEPES, 2mM glutamine, 5.0 IU/ml penicillin streptomycin (Gibco) and 10% heat inactivated Fetal Bovine Serum (FBS) (Hyclone Laboratories Inc., Logan, UT)(cRPMI).

10 *Peptide synthesis and purification and proteins*

Peptides were synthesized on a Beckman 990C peptide synthesizer as previously described (MacNeil et al., 1993). Peptides were then purified by HPLC on a reverse phase C18 column with water/acetonitrile gradient. Peptides were dissolved at 2 mg/ml in distilled H₂O and filter sterilized through a 0.22 μ m filter and further diluted in
15 either cRPMI 1640 before use in proliferation assays, or emulsified in CFA or IFA for immunization of mice. Human plasma VLDL purified apoE (Calbiochem) was used as native apoE.

Single cell suspension preparation

Mice were euthanized in a CO₂ chamber and either spleens or lymph nodes
20 were removed aseptically and immersed in ice-cold PBS. Cells were separated by mincing tissues through a fine-mesh sieve. The cells were then pelleted by centrifugation at 1500 rpm for 5 min and supernatant discarded. Erythrocytes in spleen preparations were lysed using ACK lysis buffer (0.15M NH₄Cl, 1.0 mM KHCO₃, 0.1 mM Na₂EDTA, pH 7.3) for 2 minutes at room temperature and then resuspended in PBS and pelleted again twice to wash
25 cells.

T cell proliferation assay

BALB/c mice were injected under light anesthesia with 50 μ l volumes containing 0, 1, 10 or 100 μ g/ml of apoE peptide emulsified in IFA or CFA in the hind footpad. After 10 days draining (popliteal) lymph nodes were removed and T cells were
30 purified by passage through nylon wool. 5×10^5 purified T cells and 2×10^5 gamma-irradiated (⁶⁰Co, 2500 rads)(Atomic Energy, Canada) autologous spleen APC's were incubated with 0, 1, 10, or 100 μ g/ml apoE peptide in 96 well flat bottom microtitre plates at 5% CO₂, 37°C for 3 days. PPD (20 μ g/ml) served as a positive control where CFA was used. Either APC's or T cells alone incubated with either stimulating peptide served as negative controls. 50 μ l
35 [³H]-TdR (0.5 μ Ci/well) was added for an additional 18 hours and then cells were then harvested (Tomtec, Orange, CONN). [³H]-TdR incorporation was measured on a Microbeta Liquid Scintillation Counter (Wallac, Turku, Finland).

Proliferation assays

Unprimed spleen or peritoneal exudate cells (PEC's) were incubated with 0, 1, 10, or 100 µg/ml apoEp1.B or negative control in 96 well flat bottom microtitre plates at 5% CO₂, 37°C for 2 days. 50 µl [³H]-TdR (0.5 µCi/well) was added for an additional 18 hours and then cells were then harvested (Tomtec, Orange, CONN). [³H]-TdR incorporation was measured on a Microbeta Liquid Scintillation Counter (Wallac, Turku, Finland). PU5-1.8, J77A1.4, A20, and U-937 cell lines were treated similarly, but incubated with peptide for 12 hours prior to addition of [³H]-TdR for an additional 18 hours. Results from U-937 cell lines are presented in Figure 15. Cell supernatants were harvested from most cell cultures before radioisotope addition and assayed for cytokine content.

Mixed Leukocyte Reaction Assays

Primary allogeneic MLR was set up with NOD splenocytes as stimulators and nylon wool enriched BALB/c naïve T cells as responders. Stimulator cells were pre-incubated for 3 days with different combinations of apoEp1.B, GM-CSF, and PU5-1.8 supernatant containing TNF. Stimulator cells were then treated with mitomycin C (50 µg/ml, 20 min at 37°C; Sigma) and co-cultured with 4 x 10⁴ responder for 3 days at which time cells were pulsed with 50 µl [³H]-TdR (0.5 µCi/well) for 24 hr. Cells were harvested (Tomtec) and [³H]-TdR incorporation was measured on a Microbeta Liquid Scintillation Counter (Wallac). Each bar represents the mean cpm from triplicate cultures.

Cytokine Analysis

Culture supernatants were tested for IL-2, IL-4 and IFN_γ concentration using sandwich ELISA assays. Manufacturer's (Pharmingen) protocols were followed. 96-well microtitre ELISA plates were coated with 50 µl, 1 µg/ml α-IL-2, α-IL-4 or α-IFN_γ mAB in coating buffer (0.1 M NaHCO₃, pH 8.2) overnight at 4°C. Plates were washed twice with PBS-T (PBS, Tween-20 (Sigma, St. Louis, MO) and plates were blocked for 2 hr at room temperature with 100 µl blocking buffer (PBS, 5% BSA). Plates were washed twice and 50 µl samples were added in triplicate for an overnight, 4°C incubation. Plates were then washed 5 times and following a 2 hr room temperature incubation period with 50 µl, 1 µg/ml biotinylated α-IL-2, α-IL-4 or α-IFN_γ mAB (Pharmingen), plates were washed 8 times. 50 µl, 1 µg/ml streptavidin-alkaline phosphatase (Pharmingen) was then added for 1 hr at room temperature. Following 8 washes, 50 µl p-nitrophenyl phosphate substrate (pNPP (Sigma)) was added and absorbance determined at O.D. 405 nm using a microplate reader (Biorad, Hercules, CA) after colour had developed.

Immunofluorescence and flow cytometry

1 x 10⁶ cells were washed with ice cold PBS and resuspended in 25 µl PBS and incubated with 10 µl normal mouse serum and fluorochrome conjugated anti-mouse

CD11a, CD11b, CD14, MHC class II, CD95, CD62L, CD62E, B7-1, B7-2 and ICAM-1 (Pharmingen, Mississauga, Canada) monoclonal antibodies (mAb's) for 45 minutes on ice. Dendritic cell markers DEC-205, 33D1, CD11c (N418) unconjugated antibodies were similarly treated, however, following primary antibody incubation, cells were washed 3 times and a secondary goat anti-rat (for 33D1 and DEC-205 ab's) and goat anti-hamster (for CD11c ab) fluorochrome-conjugated antibody was then added for 45 min on ice.. Cells were then washed three times. All samples were resuspended in 300 µl PBS and the fluorescence of stained cells was analyzed on a FACScan flow cytometer (Becton Dickinson, Mountain View, CA) and data collected was analyzed using Lysis II (Becton-Dickinson) software.

10 *Cell Cycle Analysis*

1 x 10⁶ cells were washed and resuspended in hypotonic propidium iodide (PI) staining solution (0.1% [w/v] sodium citrate, 0.1 [v/v] Triton X-100, 0.05 µg/ml propidium iodide) and left to stain in the dark overnight at 4°C. Cells were analyzed on a FACScan flow cytometer and the percentage of cells in each phase of the cell cycle were quantitated using ModFit software (Becton Dickinson). Aggregates were excluded from the analysis with the use of the doublet discrimination module and subsequent gating on the linear red (FL2) fluorescence area and width parameters.

Intraperitoneal Injection

BALB/c mice were injected i.p. with 300 µg apoE peptide and 48 hours later PECs were collected post-mortem by peritoneal lavage using ice cold saline.

Adoptive transfer studies

RBC-depleted diabetogenic splenocytes from 17-20 wk-old female NOD mice were resuspended in saline. 8 to 10 wk old NOD.SCID mice were injected i.p. with 0.2 µl saline containing 10⁷ of the diabetogenic splenocytes. These same mice were then immunized with either 200 µg/ml apoEp1.B or 200 µg/ml apoEp2 via the footpad. Urine glucose levels were monitored biweekly using glucose enzymatic test strips (Eli Lilly, Toronto, Ont.). Once urine tested positive, then blood glucose levels (BGL) were tested using Glucometer Encore (Miles/Bayer). Mice exhibiting BGL > 11.1 mmol/L (200 mg/dl) for 2 consecutive weeks were considered diabetic.

30 **RESULTS**

ApoEp1.B peptide induces dendritic-like cell morphology

Unprimed splenocytes, enriched spleen monocytes and monocytic PU5-1.8 and J77A4.1 cell lines from BALB/c mice were incubated with 0, 1, 10, or 100 µg/ml apoEp1.B (239-252) or negative control apoEp1.D (236-249) peptide to assess activity. After only 2 hr, apoEp1.B, but not apoEp1.D-incubated cells started to detach from plastic plates and aggregate in suspension optimally at 100 µg/ml. After 24 hr, morphologically these cells appeared less rounded, more granular. Photographs were taken after 24 hours, the results are shown in Figure 1 (magnification 400X). At 48 hours they displayed

dendritic-like processes. FACS analysis confirmed an increase in cell size and granularity (Figure 2).

ApoEp1.B peptide induces DC-like marker expression

Morphologically apoEp1.B-treated cells appeared DC-like. This was confirmed this by FACS analysis. Unprimed BALB/c splenocytes and PU5-1.8 cells were incubated for 24 hr with 100 µg/ml apoEp1.B or control peptide apoEp1.D and then stained for FACS analysis. Surface expression of CD14, CD11a, CD11b, CD11c, B7-1, B7-2, DEC-205, 33D1, MHC class II, fas, CD40, CD62L (L-selectin) and CD43 (leukosialin) were increased above background on apoEp1.B-incubated PU5-1.8 cells after 24 hr incubation (Figures 3 and 4). With repeated experimentation, expression of Mac-3 and CD54 (ICAM-1) were found to be relatively variable.

To assess whether apoEp1.B-induced phenotypic changes are dose-dependent, 10⁵ PU5-1.8 cells were incubated for 20 hr with 0 (medium), 1, 10 or 100 µg/ml apoEp1.B in 24-well plates and were then stained for FACS analysis. 10,000 events were collected. B7-1 marker is shown. The results, shown in Figure 5 illustrate that 100 µg/ml apoEp1.B was found to be optimal, however, 1 µg/ml resulted in slight phenotypic changes.

Surface expression of B7-1, B7-2, MHC class II, CD28, CD11a, CD11c and 33D1 were increased above background on apoEp1.B-incubated splenocytes after 24 hr incubation (see Figure 4). There was also an increase in CD40 and a decrease in L-selectin (data not shown) on apoEp1-treated splenocytes. There were no changes in the expression of CD16/32 at any time.

ApoEp1.B peptide-induced differentiation and activation is not strain or species-specific

Unprimed B6 and NOD splenocytes were incubated with 100 µg/ml apoEp1.B or apoEp1.D to determine whether apoEp1.B was strain-specific. After 48 hr in culture, cells were stained for FACS analysis. B6 and NOD splenocytes responded to apoEp1.B with similar marker expression increase to that of BALB/c splenocytes (data not shown). Naive spleen cells from apoE deficient (apoE K.O.) mice were also tested to determine whether in vivo production of apoE had any influence on this effect. ApoE K.O. splenocytes also responded with a similar marker expression increase to that of BALB/c (data not shown). Since human apoEp1.B (HapoEp1.B) (239-252) peptide has a high degree of homology to murine apoEp1.B (239T to 239A and 248I to 248A), species cross-reactivity was tested. Human monocytic cell line U-937 responded within 24 hr to 100 µg/ml apoEp1.B with similar, but less profound morphological and phenotypic changes to that of PU5-1.8 cells (Figure 15). To determine whether the HapoEp1.B had a similar effect on murine cells, 10⁵ PU5-1.8 cells were incubated with 100 µg/ml HapoEp1.B or murine apoEp1.B for 24 hr. The cells were then stained for FACS analysis. HapoEp1.B

induced cell clustering and an increase in similar markers to that of murine apoEp1.B, but to a lesser degree (B7-1 marker expression shown in Figure 6).

ApoEp1.B induces DC-like cells when injected intraperitoneally

To assess whether apoEp1.B could induce DC-like cells *in vivo*, BALB/c mice were injected i.p. with 300 µg/ml apoEp1.B or apoEp1.D. After 48 hr PEC's were stained for FACS analysis. ApoEp1.B induced an increase in cell size and granularity (Figure 8A) and an increase in surface expression of DEC-205, 33D1, CD11c, B7-1 and B7-2 (Figure 8B). Unlabelled peaks correspond to apoEp1.D. Mean fluorescent intensity shown in brackets. The results demonstrate that apoEp1.B induces differentiation and activation of PECs *in vivo* similar to the *in vitro* system.

To assess functional activation of apoEp1.B treated cells, BALB/c unprimed splenocytes were incubated with or without apoEp1.B (100µg/ml) *in vitro* for 24 hours. Cells (washed and unwashed) were stimulated with ConA for 12 hours and 3H was then added for an additional 12 hours. Cells were then harvested and radioactivity uptake counted. Results are presented as the mean of triplicate wells (cpm+/- SEM). As shown in Figure 9, apoEp1.B treated cells respond to mitogen 3-fold higher than untreated cells.

Cytokines produced after apoEp1.B immunization

ApoEp1.B or apoEp1.D-immunized NOD lymph node cells were cultured for 48 hr in the absence of challenge antigen or exogenous cytokine. Cell supernatants were harvested and tested for cytokine content to assess whether apoEp1.B induced a Th1 or Th2 response. ApoEp1.B-primed cells secreted higher levels of IL-4 and lower levels of IL-2 than apoEp1.D primed cells (Figure 12). There was no change in IFNγ.

T cell allostimulatory activity of apoEp1.B-induced DC

To measure the T cell allostimulatory or inhibitory ability of these DC's, 4 x 10⁴ enriched BALB/c naïve T cells were co-cultured with 40, 400, 4000 and 40,000 mitomycin C-treated NOD splenocytes which had been incubated with various stimulants for 3 d. T cell proliferation was measured following 3 days of co-culture by [³H]-TdR incorporation. The results are shown in Figure 12. Bars represent the mean of triplicate wells (cpm +/- S.D.). * and ** represent p<0.05 and p<0.001, respectively in 2-way ANOVA's compared to medium alone (nil) controls. ApoEp1.B-treated APC allostimulation was similar to that of untreated APC's at 4 x 10⁴ APC's per well concentration, however, there was a slightly higher level of apoEp1.B-treated APC-T cell stimulation over untreated APC's at 4 x 10³ and 4 x 10² APC's. There was a dramatic increase in allostimulation when APC's were pre-incubated with GM-CSF, particularly at 4 x 10³ APC's per well. This increase was slightly, but significantly reduced when APC's were pre-incubated with GM-CSF plus apoEp1.B. The more striking result is depicted in Figure 5B. APC's (4 x 10³) in all bars (except "T alone") have been untreated (nil) or

pre-incubated with 100 µg/ml apoEp1.B, apoEp1.B + GM-CSF, or GM-CSF alone for 3 days prior to co-culture. Upon combining responder T cells with APC, various exogenous factors were added, including 20 µg/ml Con A and 5 µg/ml apoEp1.B. Very little proliferation was detected in wells containing T cells alone or APC alone. T cells proliferated well in response to Con A, however, when apoEp1.B was added, T cell allostimulation was completely abrogated.

DISCUSSION

It is known that monocytic cells can differentiate into DC's, however, GM-CSF has been long been thought to be essential for this effect (Scheicher *et al.*, 1992). It has been shown here that a novel self peptide, apoEp1.B, alone induces DC's *in vitro* from both naive monocytic cells, and a murine cell line. These DC's possess the morphology and cell surface markers of classical DC's. ApoEp1.B also induces DC's *in vivo* upon apoEp1.B immunization. These DC's may favour a Th2 response as shown by an increase in IL-4 and a decrease in IL-2 production, and may therefore, be involved in split tolerance.

ApoEp1.B induced DC morphology of naive BALB/c splenocytes, enriched spleen monocytes (data not shown), monocytic cell lines PU5-1.8 (Figure 1) and J77A4.1 *in vitro* (data not shown). Following only 2 hr incubation, apoEp1.B-treated cells started to detach from plastic plates and aggregated in suspension, a process characteristic of DC's. Since there was increased cell death, proliferation assays did not show an overall increase in cell number (data not shown). This is consistent with the loss of proliferation characteristic of cell differentiation.

ApoEp1.B-treatment of PU5-1.8 cells induced an increase in DC-specific markers, DEC-205, 33D1 and CD11c. Other markers that consistently increased were CD11a, B7-1, B7-2, MHC class II, fas, CD40 and L-selectin.

ApoEp1.B stimulation of splenocytes induced an increase in CD40, CD11a, CD11c, 33D1, B7-1, B7-2, MHC class II, fas and fasL expression at 24 hr. A similar profile was recorded following 72 hr post-apoEp1.B treatment (data not shown). These changes were not as dramatic as apoEp1.B-induced PU5-1.8 surface marker changes. This does not detract from the results obtained with PU5-1.8 cells as these cells are transformed and capable of rapid growth, cell division and protein synthesis.

Changes in B7 costimulatory molecules would inevitably alter T cell responses. Furthermore, an increase in MHC class II enables DC's to present peptides at a higher density which renders them more efficient at either T cell activation or tolerance. An increase in expression of adhesion molecules such as LFA-1 may enable DC's migration from various tissues to lymphoid organs where they present captured Ag.

In contrast to splenocytes, L-selectin expression on PU5-1.8 cells increases with apoEp1.B incubation (data not shown). L-selectin is rapidly downregulated on activated cells and thus the spleen cell data suggests that apoEp1.B activates these cells.

These conflicting results are not extraordinary when considering that PU5-1.8 cells are transformed as well as a homogenous population. Heterogeneous cell-cell interactions as well as paracrine stimulatory and inhibitory factors that affect immune responses may explain some of the differences between spleen and PU5-1.8 cells. Furthermore, transformed
5 PU5-1.8 cells may lack mechanisms necessary for L-selectin downregulation, such as specific proteases for L-selectin cleavage. CD28 expression on splenocytes is also increased slightly with apoEp1.B treatment.

A slight increase in macrophage differentiation markers CD14 and Mac-3, indicates that maturation of macrophages may be also occurring in apoEp1.B-treated
10 cultures. Whether these cells then develop into DC's, apoptose or remain as macrophages is unknown.

Following i.p. injection, apoEp1.B induced an increase in PEC size and granularity and an increase in DC-specific markers, DEC-205, 33D1 and CD11c as well as B7-1 and B7-2. Therefore, injection of apoEp1.B i.p. induces PEC's to express a similar
15 DC-like phenotype compared to *in vitro* experiments. The increase in DC marker expression is not as pronounced as it was *in vitro*, probably due to the dilution and/or clearance of apoEp1.B peptide *in vivo*. However, since DC's are efficient at T cell stimulation or inhibition, even a modest increase in number may have a profound effect on an immune response.

20 The treatment of NOD mice with apoEp1.B stimulated an increase in IL-4 secretion, presumably by Th2-like cells and a decrease in IL-2. The immunization of mice with apoEp1.B also induced an increase in DC's.

Whether apoEp1.B is a naturally cleaved product of apoE is unclear but doubtful. ApoEp1.B binds I-A^d with a similar affinity to that of the original apoEp1
25 peptide (which does not induce DC's), yet was not eluted in elution experiments, it is therefore, probably not naturally cleaved a high levels.

Here we show that apoEp1.B, stimulates mo/ma activation. In addition, apoEp1.B induces DC's that may favour a Th2-like response. These DC's may be partially activated or not fully mature, fully activated or induced to full maturity by inflammatory
30 signals.

Example 2

ApoEp1.B Inhibits Atherosclerotic Plaque Development

To investigate the role of apoEp1.B peptide in inflammation, particularly in response to arterial injury, a balloon angioplasty injury model which has been well
35 characterized by Dr. Alex Lucas (Lucas et al. 1996 Circ. 94:2898-900) was used. 300 µg/ml of apoEp1 or apoEp1.B peptide were infused intra-arterially immediately prior to angioplasty at the site of injury in the iliofemoral artery of the rat. Positive control SERP-1 and negative control saline alone were also used. The rats were monitored carefully and

euthanized 6 weeks after treatment and iliofemoral arteries were assessed for plaque development by histochemistry. Careful measurement of arterial thickness and lumen size showed apoEp1.B (500 μ g) peptide reduced lesion size in this model. In fact, in many of the apoEp1.B treated rats, no plaque was detected. No adverse effects were observed in apoEp1.B treated rats. The results are presented in Table 3 and Figure 10.

Example 3

ApoEp1.B immunization protects mice from diabetes

8 female 8 week old NOD mice were immunized with 250 μ g apoEp1.B peptide emulsed in IFA in one footpad. 6 negative control mice were immunized with saline in IFA. Mice were monitored for a following 4 months and their urine glucose tested to assess diabetes development. At the 8 month end date, all 8 apoEp1.B immunized mice still remain diabetes free. 4 of the 6 mice treated with saline in IFA are deceased due to diabetes. The results, shown in Figure 14, demonstrate that apoEp1.B immunization protects diabetes prone mice.

Example 4

ApoEp1.B-immunization induces Th2-like cells

NOD mice were immunized via the footpad with 200 μ g/ml apoEp1.B or apoEp1.D peptide in IFA. Draining lymph nodes were removed following 7 days and cells cultured in 96-well microtiter plates in the absence of further stimulation. Cell culture supernatants were harvested at 48 hr and tested for cytokine content. The results, shown in Figure 11, demonstrate that apoEp1.B immunization induce Th2-like cells. (Results of IL-2, IL-4 and IFN γ are presented as the mean of triplicate wells (O.D. +/- SEM). * represents $p < 0.001$ in 2-way ANOVA's compared to apoEp1.D-treated controls.)

To determine whether the Th2 response induced by apoEp1.B immunization influenced IDDM incidence, 10 female 8 week old NOD mice were immunized with 200 μ g/ml apoEp1.B and 10 mice with negative control apoEp2 emulsed in IFA in one footpad. Mice were monitored for the following 8 months and their urine and blood glucose tested for disease onset. At 10 months, 7 of 10 apoEp1.B-immunized mice remain diabetes free, whereas, 1 of 10 apoEp2-treated mice remains disease-free. (Figure 13A).

Example 5

ApoEp1.B immunization protects NOD.SCID mice from adoptively transferred diabetes

To test the protective effect of apoEp1.B in an alternative model of IDDM, 10⁷ diabetogenic splenocytes were adoptively transferred to 20 NOD.SCID mice. 10 of these mice were immunized with 200 μ g/ml apoEp1.B, the other with 200 μ g/ml apoEp2, the other with 200 μ g/ml apoEp2. All 10 negative controls have succumbed to disease after 5 weeks. 5 of the 10 apoEp1.B mice remain diabetes-free at 9 weeks (Figure 13B).

Discussion of Results from Examples 3, 4 and 5

Since a Th2 response can be protective in the NOD model of IDDM and since apoEp1.B immunization of NOD mice favours a Th2 type response (see Figure 11), apoEp1.B was tested for disease protection in these mice. ApoEp1.B/IFA immunization significantly
5 delayed disease onset in NOD mice and in adoptively-transferred NOD.Scid mice. ApoEp1.B-protected mice had slightly less islet infiltration as compared to unprotected mice (data not shown), indicating that apoEp1.B protection may involve the induction of regulatory cells rather than deletion of destructive effector cells.

APC's from NOD mice are thought to have functional and/or
10 differentiation defects (Serreze *et al.*, 1988, Serreze *et al.*, 1993). Since NOD Th2 cells are more dependent on B7 costimulation than Th1 cells (Rulifson *et al.*, 1997), a proinflammatory Th1 response may predominate in these mice by default. Consistent with this are studies demonstrating that Th2 cell hyporesponsiveness intrinsic to NOD mice (Zipris 1991a and 1991b) is reversible either by IL-4 (Rapoport *et al.*, 1993, Mueller *et al.*,
15 1996, Cameron *et al.*, 1997) or CD28 (Arreaza *et al.*, 1997) administration *in vivo*. Anti-CD28 treatment is thought to promote Th2 survival and expansion. Furthermore, costimulation deficient mice (CD28^{-/-}) display more severe IDDM (Rulifson *et al.*, 1997).

It is unclear what role DC's play in IDDM, however, the importance of DC's in disease is emphasized by the DC K.O. mouse (RelB^{-/-}). These mice display
20 aggressive multi-organ infiltration and inflammation. Another group reported that pancreatic DC's can induce disease protection upon transfer (Clare-Salzler *et al.*, 1992). These findings suggest that DC's may play a protective role in IDDM. Furthermore, Voorbij *et al.* (1989) showed that DC's were the first cells to accumulate around pancreatic islets in the spontaneously diabetic BB rat model, followed by lymphocytes.

Whether the induction of DC's upon apoEp1.B injection has a direct effect
25 on disease protection is unknown, however, the resulting Th2 response may be mediating protection. It is proposed that apoEp1.B immunization stimulates DC differentiation with an increase in B7 costimulatory molecules. These more efficient APC's, capable of stronger costimulation, rescue IL-4 producing Th2 cell responsiveness in diabetes-prone mice. This in
30 turn reduces disease incidence. Alternatively or additionally, apoEp1.B may act directly on T cells, increasing CD28 expression and/or lowering the threshold for stimulation. Since Th1 cells are already stimulated in NOD mice, protective Th2 or Th3 cell stimulation may be preferentially restored. Consistent with this statement is the data showing no change in IFN production, yet an increase in IL-4.

35 Since the lack of Th2 cell responsiveness intrinsic to NOD mice (Zipris 1991a and 1991b) is reversible, it is proposed the apoEp1.B immunization rescues Th2 responsiveness in diabetes-prone mice, which in turn reduces disease incidence. Since T cells require APC's for stimulation, we speculate that apoEp1.B stimulates a Th2-inducing APC,

possible of the DC phenotype. Irrespective of mechanism, apoEp1.B may offer a potential self peptide therapy for diabetes as well as other Th1-mediated autoimmune diseases.

Example 6

Modifications to ApoEp1.B Sequence

5 Various amino acid substitutions and elongations were made to the apoEp1.B peptide as illustrated in Tables 1 and 2. The modified peptides were incubated at 100µg/ml with 10⁵ PU5-1.8 cells for 20 hours. The cells were stained with B7-1 for FACS analysis. The results demonstrate that some, but not all, single a.a. substitutions, elongations or deletions can decrease or abrogate apoEp1.B effects (see Table 1 and 2 and
10 Figure 16 and 18). This supports the notion that there is a receptor to which apoEp1.B binds and certain a.a. changes decrease peptide-receptor affinity. The deletion of 239^T results in a greatly reduced effect on surface marker changes. However, the substitution of this same a.a. with an alanine (as in the sequence of HapoEp1.B) has only a slightly reduced effect compared to apoEp1.B. Therefore, the length of apoEp1.B is likely important in receptor
15 binding. It also appears that the structural integrity resides in the carboxy terminal region of the apoEp1.B peptide as even slight changes in that region decreases or abolishes the activity of the peptide.

While the present invention has been described with reference to what are presently considered to be the preferred examples, it is to be understood that the invention
20 is not limited to the disclosed examples. To the contrary, the invention is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the appended claims.

All publications, patents and patent applications are herein incorporated by reference in their entirety to the same extent as if each individual publication, patent or
25 patent application was specifically and individually indicated to be incorporated by reference in its entirety.

TABLE 1
ApoEp1.B Elongations and Truncations

	<u>Name</u>	<u>Sequence</u>	<u>Residues</u>	<u>DC Marker Changes</u>
	apoEp1	EEQ TQQ IRL QAE IFQ AR	236-252	slight
5	apoEp1.Ba	EQ TQQ IRL QAE IFQ AR	237-252	-
	apoEp1.Bb	Q TQQ IRL QAE IFQ AR	238-252	++
	apoEp1.B	TQQ IRL QAE IFQ AR	239-252	+++++
	apoEp1.Bc	QQ IRL QAE IFQ AR	240-252	++
	apoEp1.Bd	Q IRL QAE IFQ AR	241-252	-

TABLE 2
ApoEp1.B Amino Acid Substitutions

	<u>Name</u>	<u>Sequence</u>	<u>Residues</u>	<u>DC Marker Changes</u>
	apoEp1.B A ²⁴⁰	TΔQ IRL QAE IFQ AR	239-252	+++
5	apoEp1.B A ²⁴¹	TQΔ IRL QAE IFQ AR	239-252	+++++
	apoEp1.B A ²⁴²	TQQ ΔRL QAE IFQ AR	239-252	++
	apoEp1.B A ²⁴³	TQQ IΔL QAE IFQ AR	239-252	+++
	apoEp1.B A ²⁴⁴	TQQ IRΔ QAE IFQ AR	239-252	-
	apoEp1.B A ²⁴⁵	TQQ IRL ΔAE IFQ AR	239-252	-
10	apoEp1.B A ²⁴⁷	TQQ IRL QAΔ IFQ AR	239-252	-
	apoEp1.B A ²⁴⁸	TQQ IRL QAE ΔFQ AR	239-252	-
	apoEp1.B A ²⁴⁹	TQQ IRL QAE IΔQ AR	239-252	-
	apoEp1.B A ²⁵⁰	TQQ IRL QAE IFΔ AR	239-252	-
	apoEp1.B A ^{239,240}	ΔΔQ IRL QAE IFQ AR	239-252	-
15	apoEp1.B A ^{240,241}	TΔΔ IRL QAE IFQ AR	239-252	-
	apoEp1.B A ²³⁹⁻²⁴¹	ΔΔΔ IRL QAE IFQ AR	239-252	-
	human apoEp1.B	AQQ IRL QAE ΔFQ AR	239-252	++++

TABLE 3

	Lumen Area (mean +/- S.E.)	Plaque Thickness (mean +/- S.E.)	Plaque area (mean +/- S.E.)
saline	0.027 +/- 0.012	0.139 +/- 0.028	0.059 +/- 0.010
apoEpl.B	0.156 +/- 0.020	0.003 +/- 0.003	0.002 +/- 0.002
P-value	<.0001	<.0001	<.0001

FULL CITATIONS FOR REFERENCES REFERRED TO IN THE SPECIFICATION

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We Claim:

1. A method of immune modulation comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or animal in need thereof.
- 5 2. A method according to claim 1 wherein the immune modulation is the induction of cell differentiation.
3. A method according to claim 2 wherein the cell is a dendritic cell.
4. A method according to claim 2 wherein the cell is a tumor cell.
5. A method according to claim 1 wherein the immune modulation is the
10 induction of immune tolerance.
6. A method according to claim 1 wherein the immune modulation is the inhibition or prevention of inflammation.
7. A method according to claim 6 for inhibiting atherosclerotic plaque formation.
- 15 8. A method according to claim 1 wherein the immune modulation is the treatment or prevention of an autoimmune disease.
9. A method according to claim 8 wherein the autoimmune disease is diabetes.
10. A method according to claim 1 wherein the immune modulation is the induction of response.
- 20 11. A method of inducing tolerogenic dendritic cells comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to a cell or animal in need thereof.
12. A method according to any one of claims 1 to 11 wherein said peptide is administered to a cell *in vitro*.

13. A method of treating a tumor comprising administering an effective amount of an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide to an animal in need thereof.
14. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence TQQIRLQAEIFQAR (SEQ.ID.NO.:1), or a fragment, elongation, analog or derivative of the peptide.
15. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence AQQIRLQAEAFQAR (SEQ.ID.NO.:2), or a fragment, elongation, analog or derivative of the peptide.
- 10 16. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence TAQIRLQAEIFQAR (SEQ.ID.NO.:3).
17. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence TQAIRLQAEIFQAR (SEQ.ID.NO.:4).
18. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence TQQARLQAEIFQAR (SEQ.ID.NO.:5).
- 15 19. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence TQQIALQAEIFQAR (SEQ.ID.NO.:6).
20. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence QTQQIRLQAEIFQAR (SEQ.ID.NO.:7).
- 20 21. A method according to any one of claims 1 to 13 wherein the apoEp1.B peptide has the amino acid sequence QQIRLQAEIFQAR (SEQ.ID.NO.:8).
22. A method according to claim 10 further comprising administering a cytokine to the cell or animal in need thereof.
23. A pharmaceutical composition for modulating an immune response comprising an apoEp1.B peptide or a nucleic acid encoding an apoEp1.B peptide in admixture with a suitable diluent or carrier.
- 25

24. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence TQQIRLQAEIFQAR (SEQ.ID.NO.:1), or a fragment, elongation, analog or derivative of the peptide.
25. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence AQQIRLQAEAFQAR (SEQ.ID.NO.:2), or a fragment, elongation, analog or derivative of the peptide.
26. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence TAQIRLQAEIFQAR (SEQ.ID.NO.:3).
27. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence TQAIRLQAEIFQAR (SEQ.ID.NO.:4).
28. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence TQQARLQAEIFQAR (SEQ.ID.NO.:5).
29. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence TQQIALQAEIFQAR (SEQ.ID.NO.:6).
30. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence QTQQIRLQAEIFQAR (SEQ.ID.NO.:7).
31. A pharmaceutical composition according to claim 23 wherein the apoEp1.B peptide has the amino acid sequence QQIRLQAEIFQAR (SEQ.ID.NO.:8).
32. A pharmaceutical composition according to claim 23 further comprising a cytokine.
33. An isolated apoEp1.B peptide which includes amino acids 239-252 of the apolipoprotein E protein, or an analog, fragment, elongation or derivative of the peptide.
34. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence TQQIRLQAEIFQAR (SEQ.ID.NO.:1) or an analog, fragment, elongation or derivative of the peptide.

35. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence AQQIRLQAEAFQAR (SEQ.ID.NO.:2) or an analog, fragment, elongation or derivative of the peptide.

36. An isolated apoEp1.B peptide according to claim 33 having the amino acid
5 sequence TAQIRLQAEIFQAR (SEQ.ID.NO.:3).

37. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence TQAIRLQAEIFQAR (SEQ.ID.NO.:4).

38. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence TQQARLQAEIFQAR (SEQ.ID.NO.:5).

10 39. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence TQQIALQAEIFQAR (SEQ.ID.NO.:6).

40. An isolated apoEp1.B peptide according to claim 33 having the amino acid sequence QTQQIRLQAEIFQAR (SEQ.ID.NO.:7).

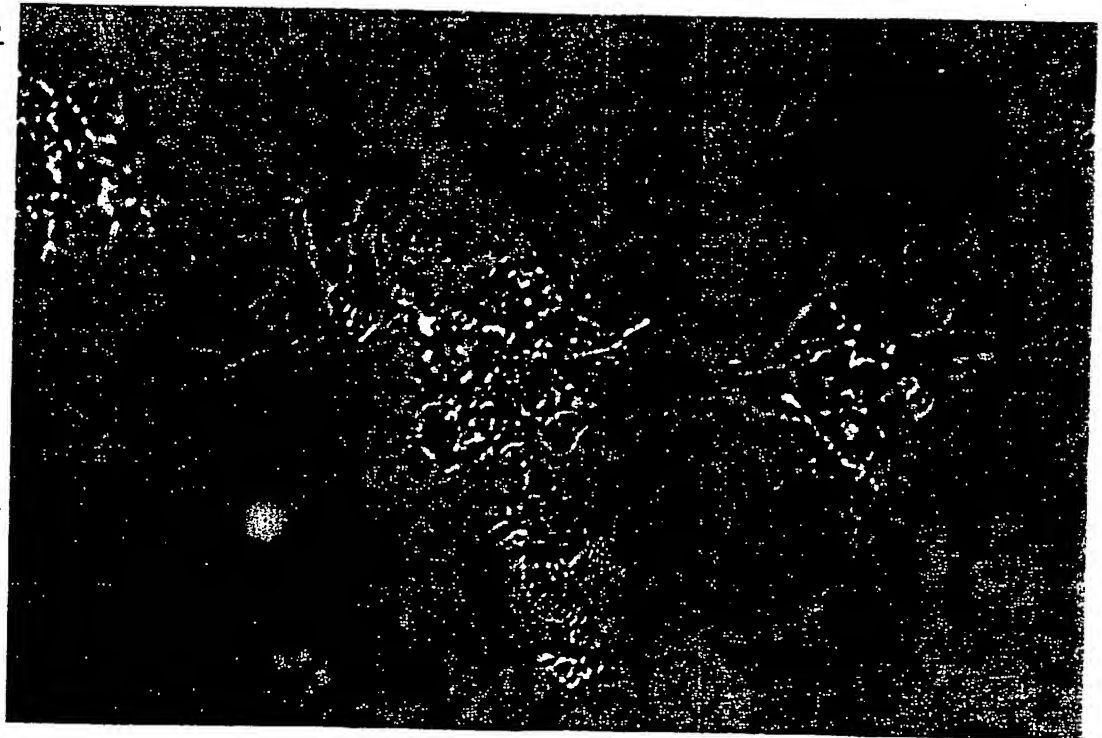
41. An isolated apoEp1.B peptide according to claim 33 having the amino acid
15 sequence QQIRLQAEIFQAR (SEQ.ID.NO.:8).

1/20
FIGURE 1

A

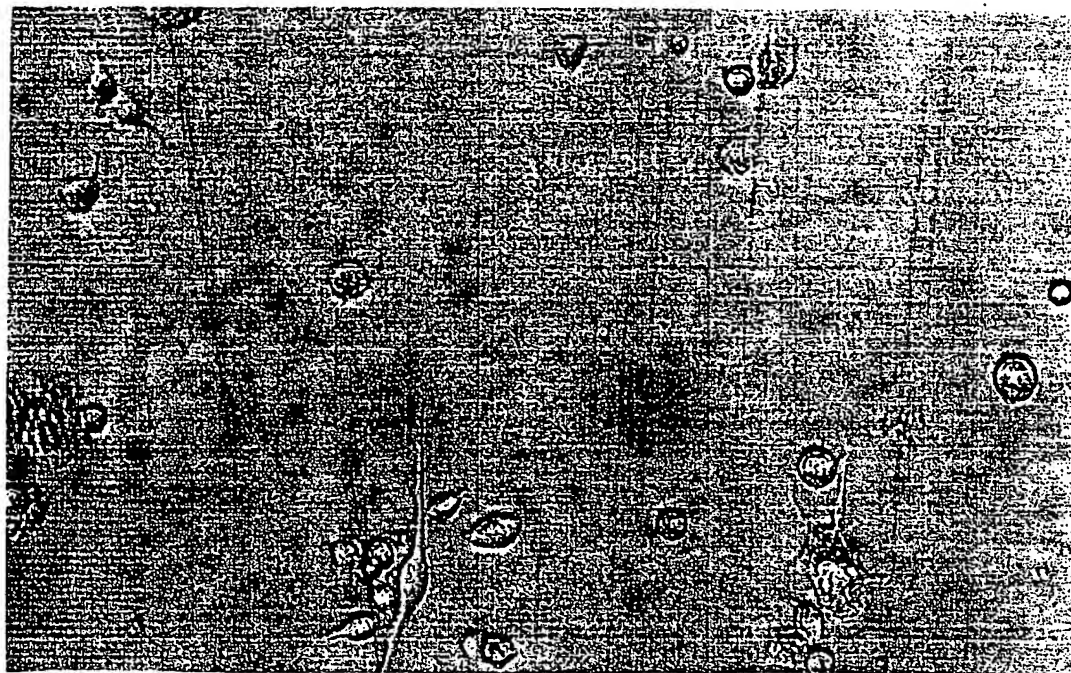


B

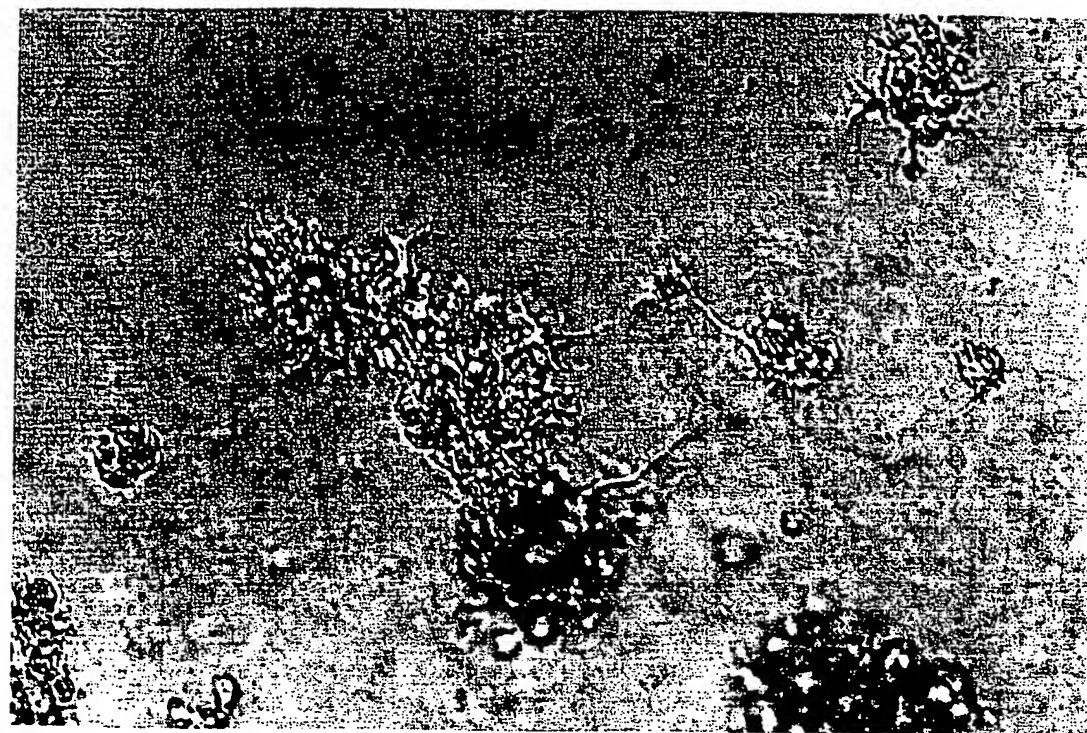


2/20
FIGURE 1 cont'd

C

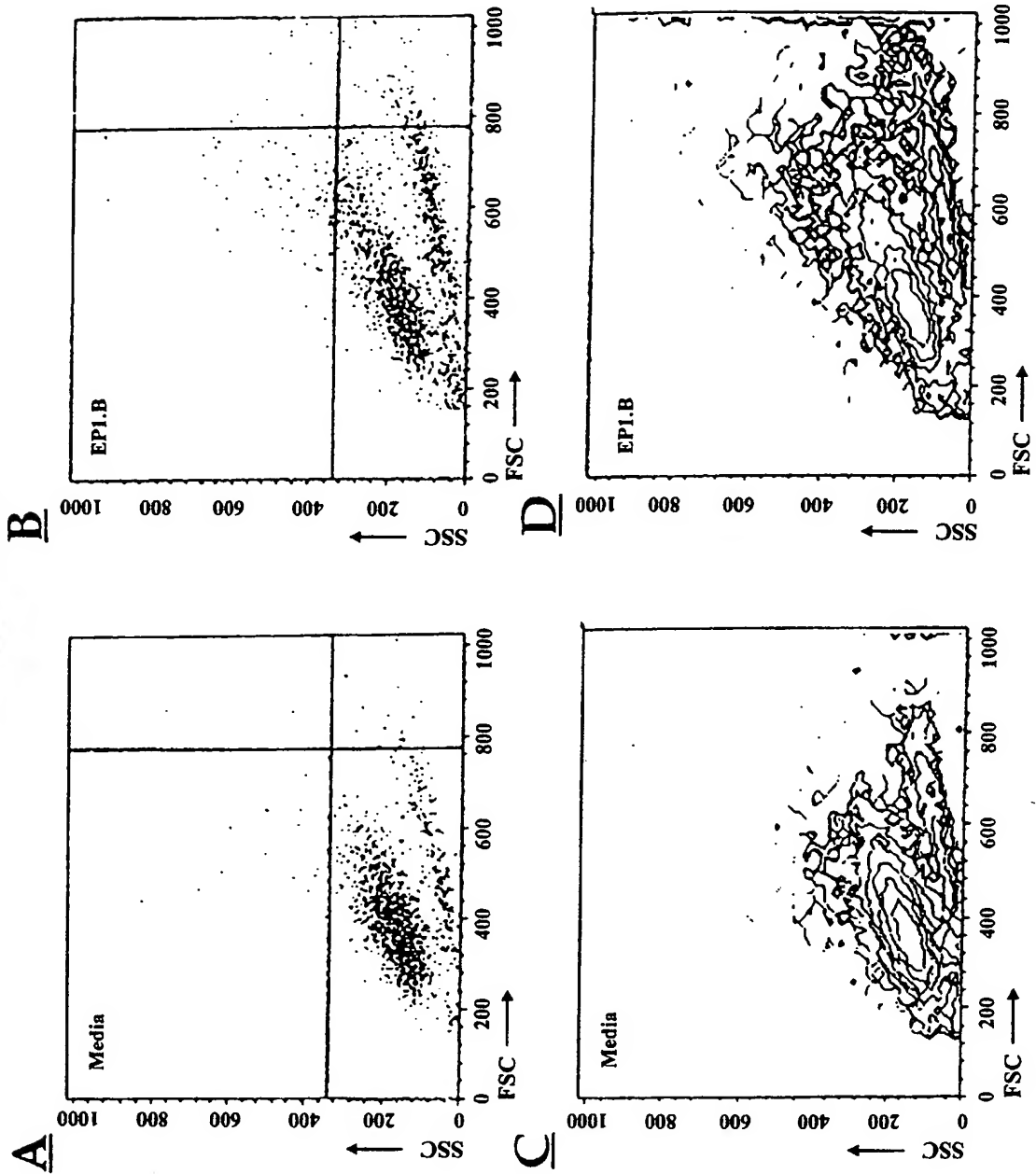


D

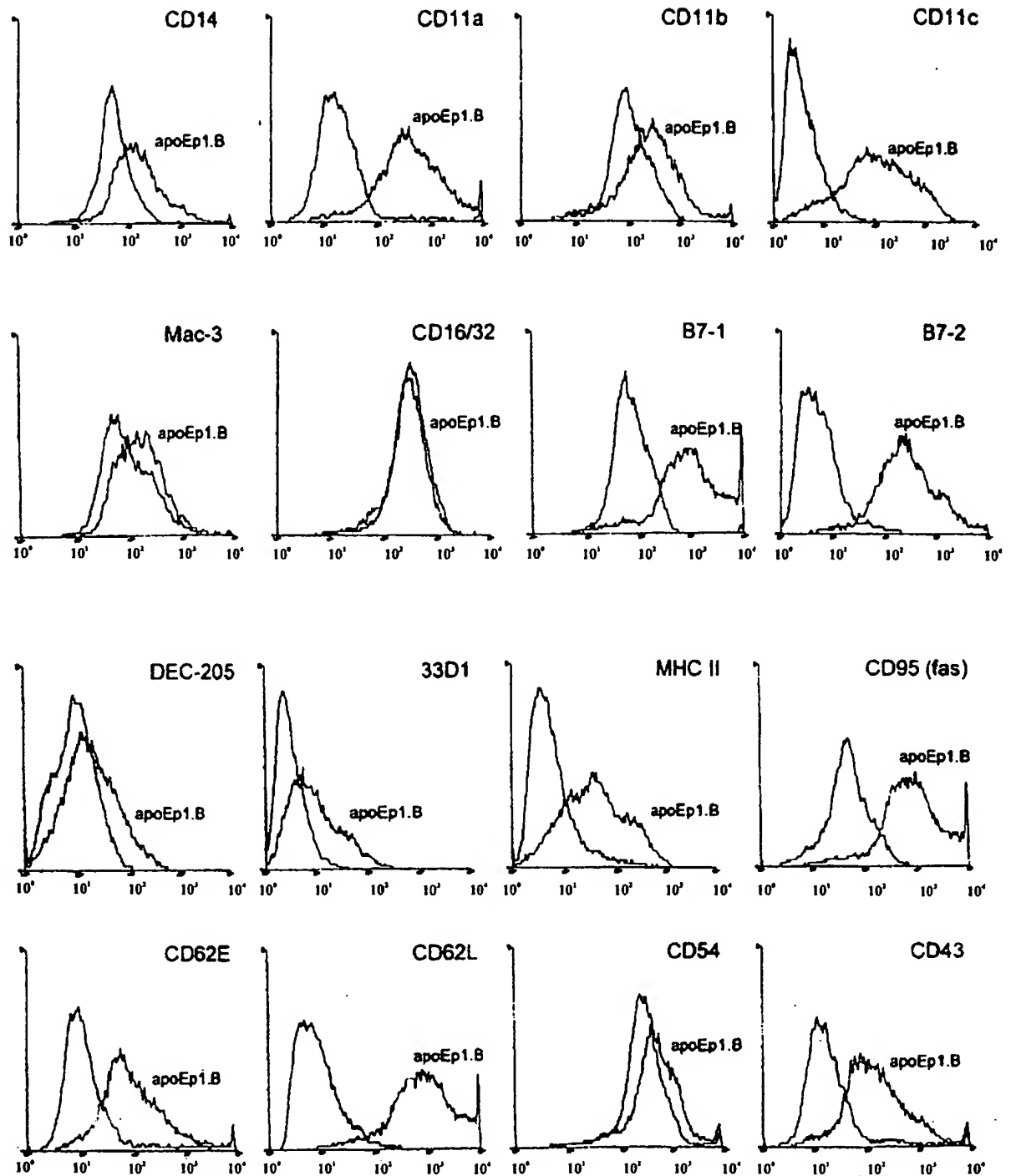


3/20

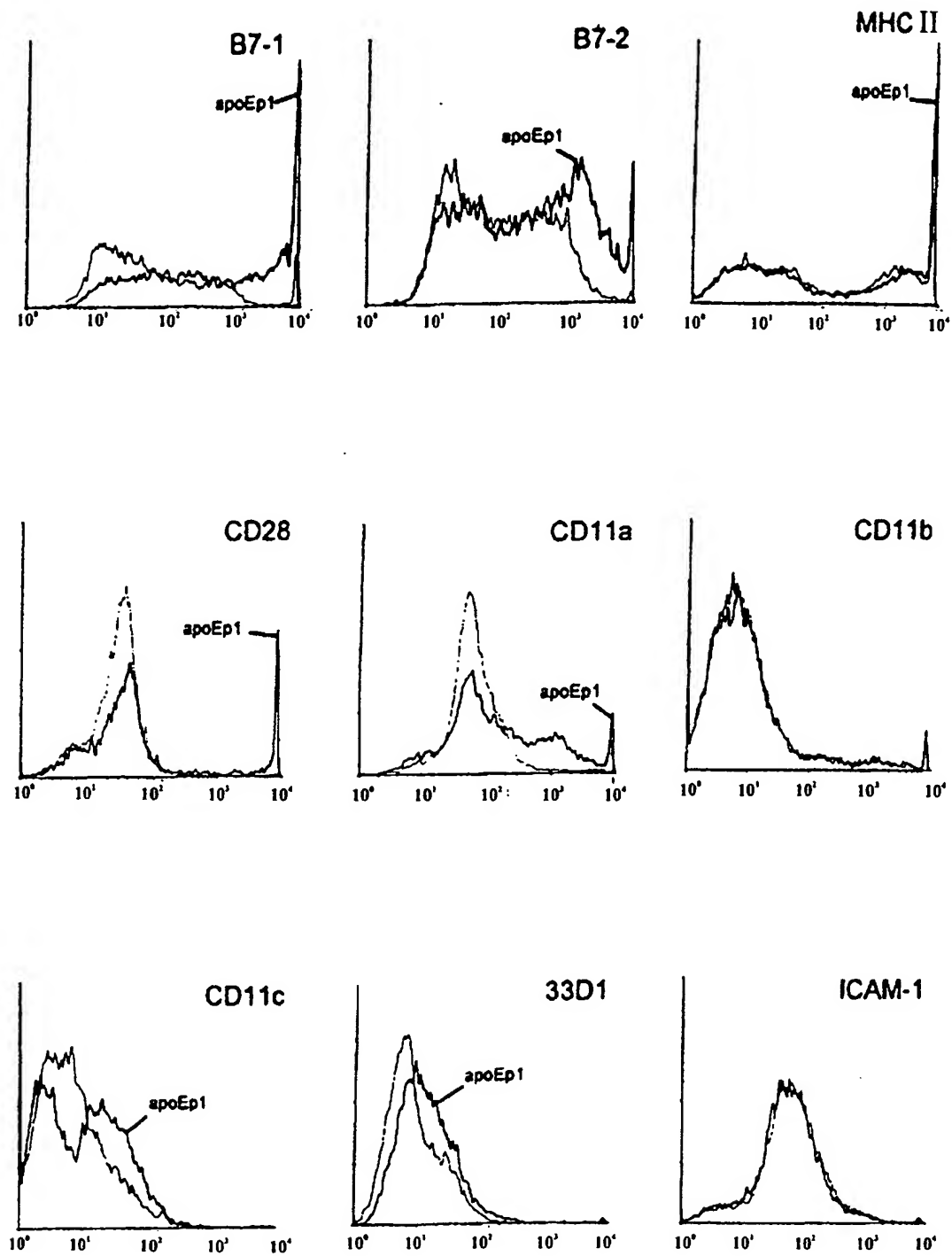
FIGURE 2



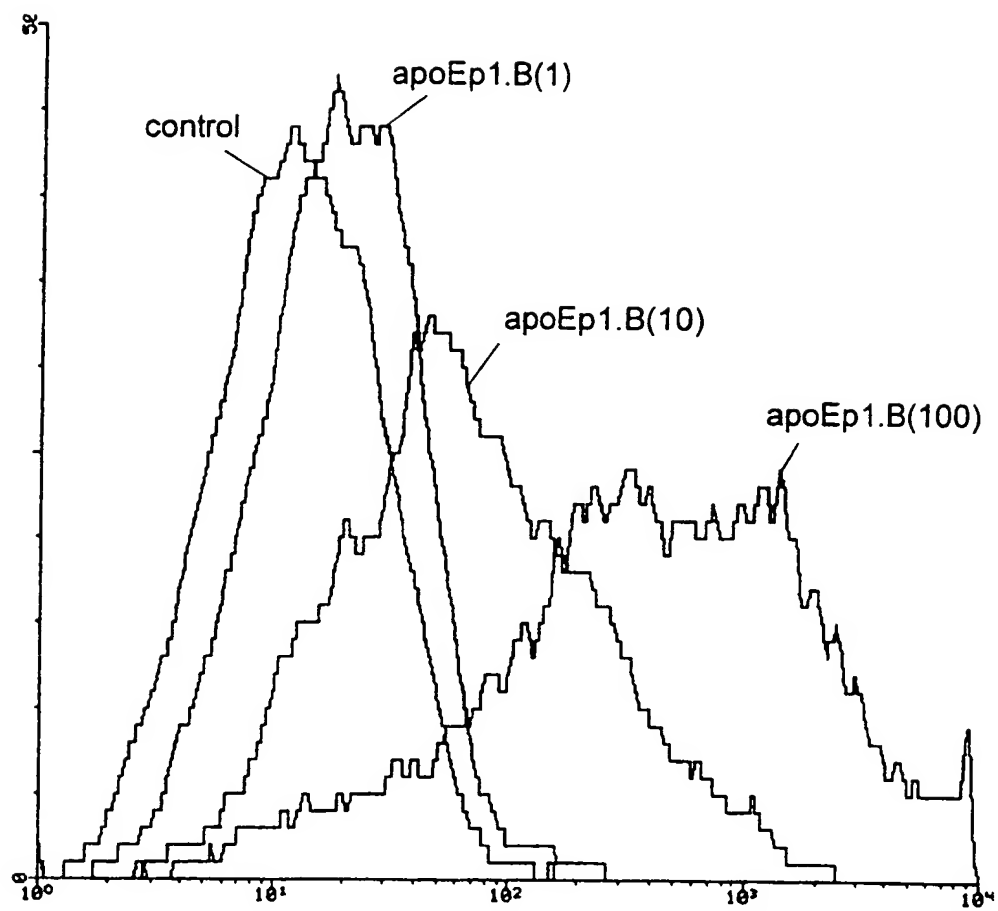
4/20
FIGURE 3



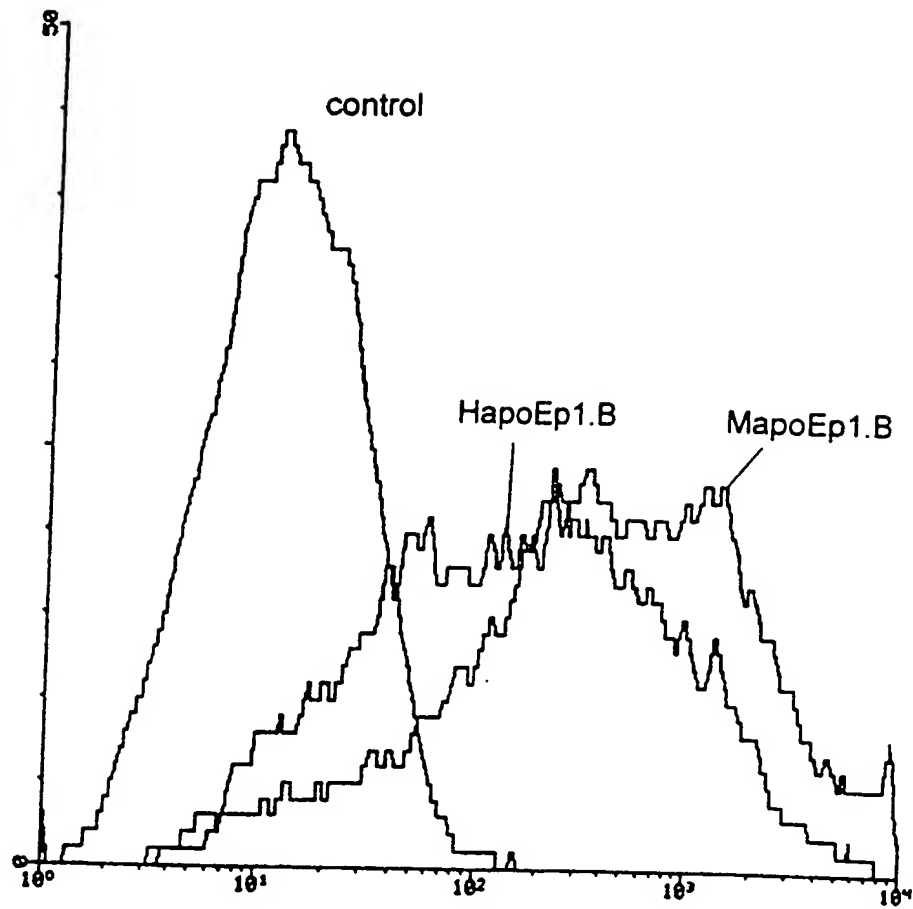
5/20
FIGURE 4



6/20
FIGURE 5

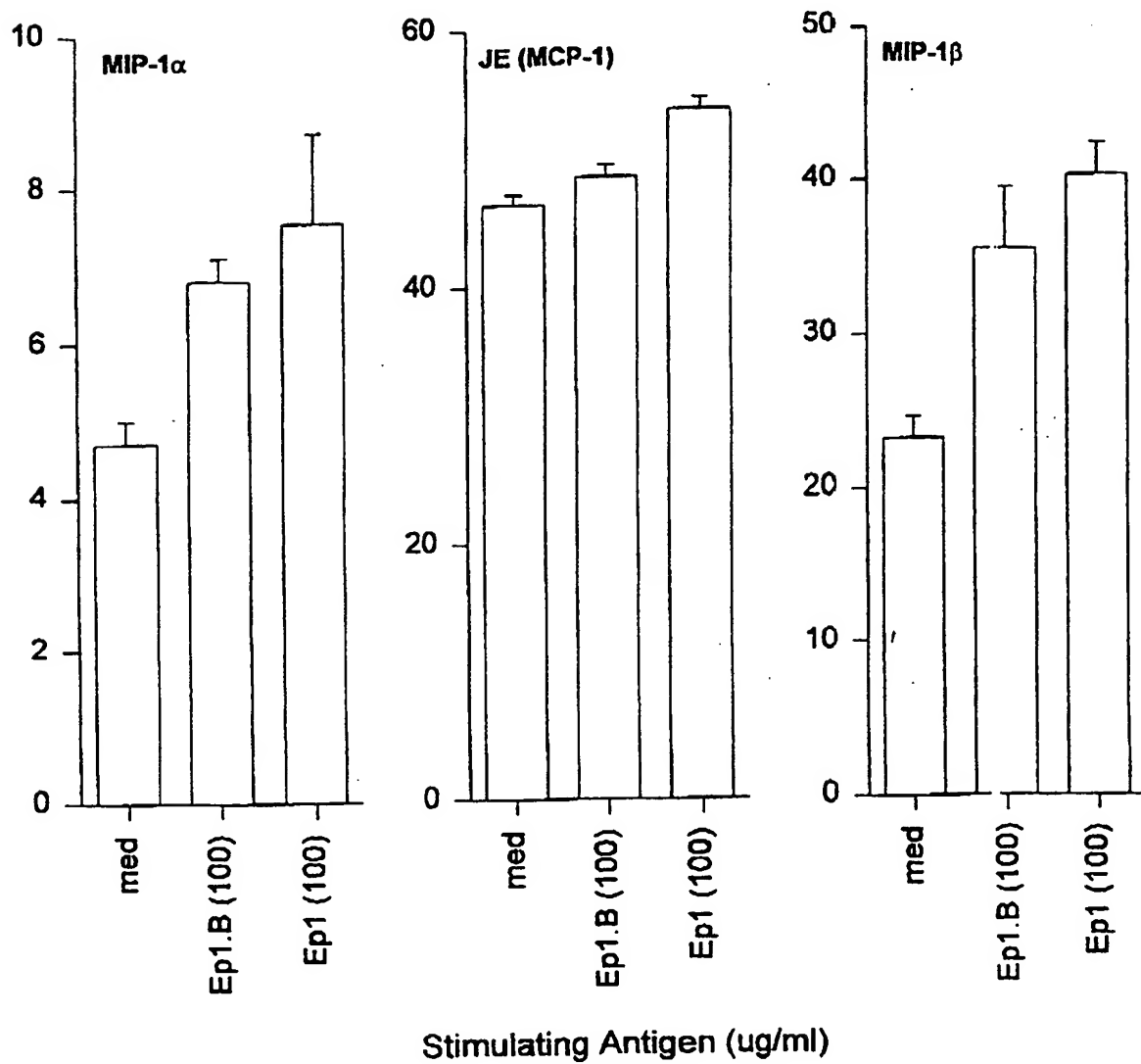


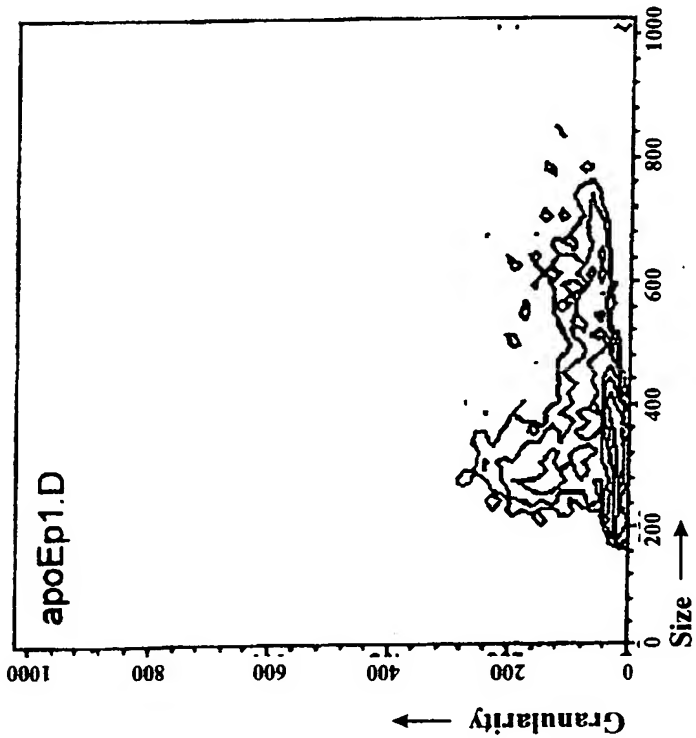
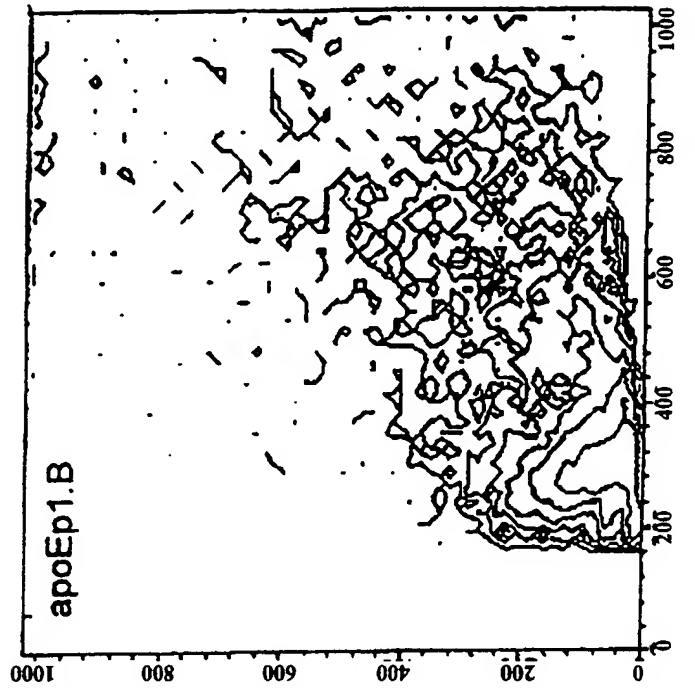
7/20
FIGURE 6



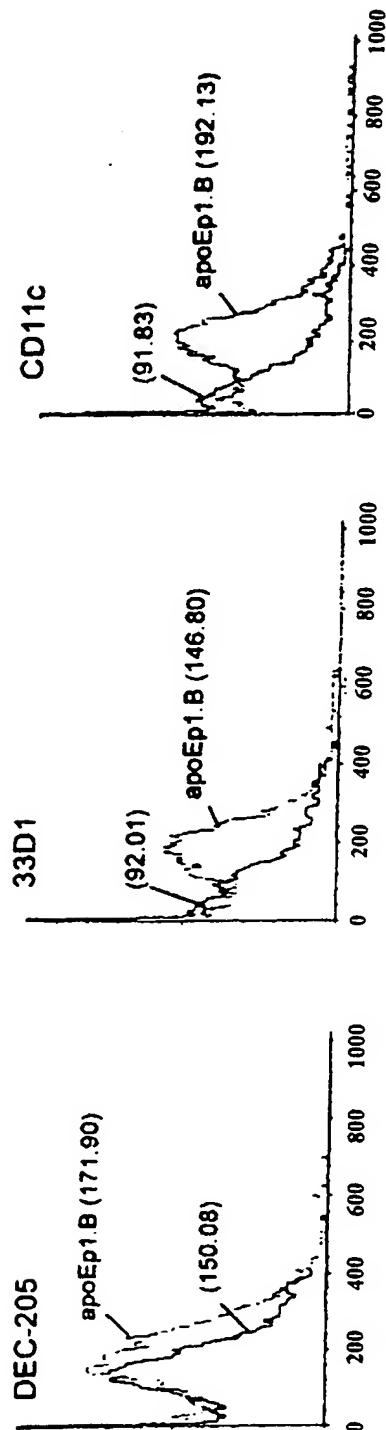
8/20
FIGURE 7

Level of chemokine in 24 h supernatant



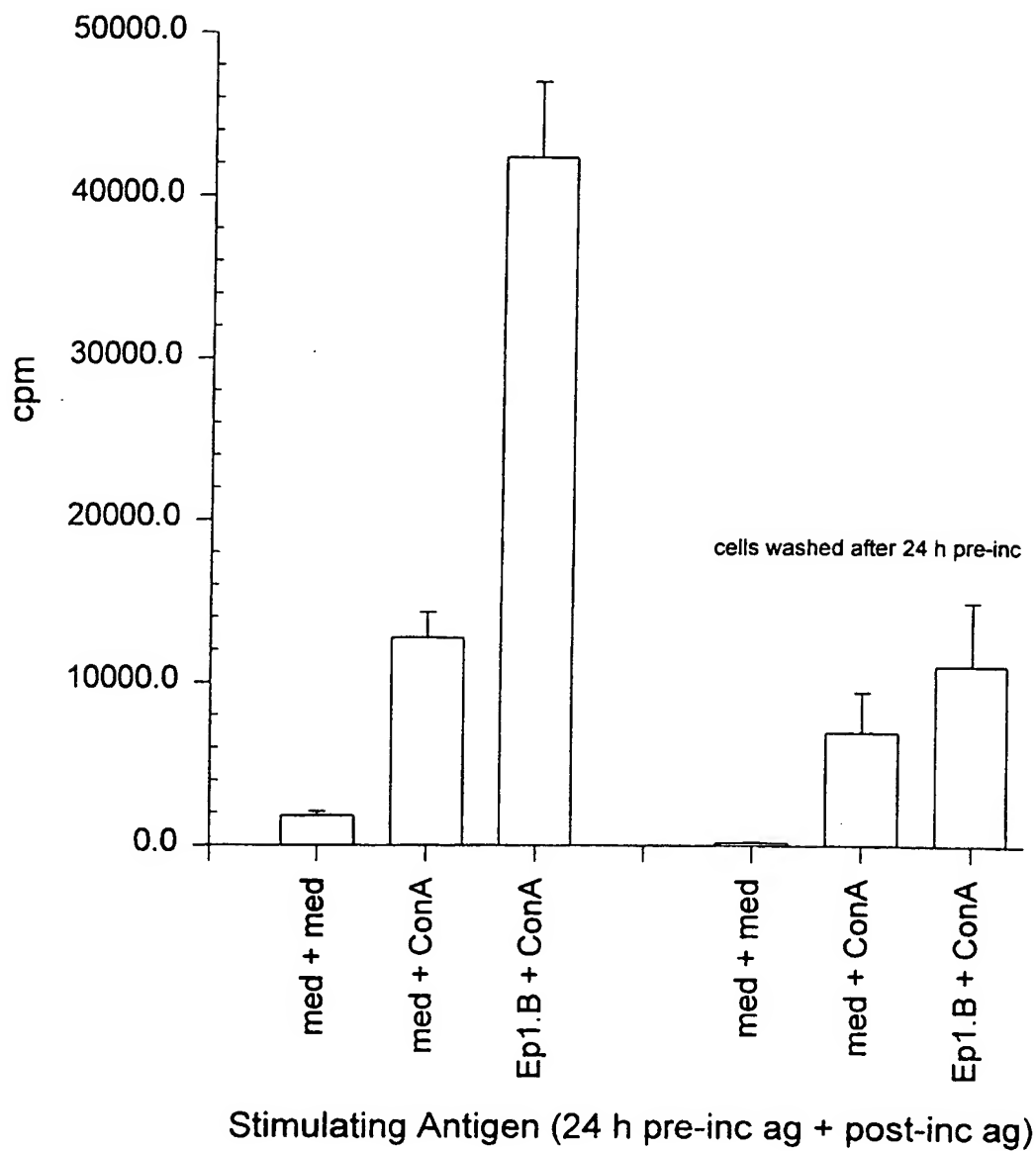
9/20FIGURE 8A

SUBSTITUTE SHEET (RULE 26)

10/20FIGURE 8B

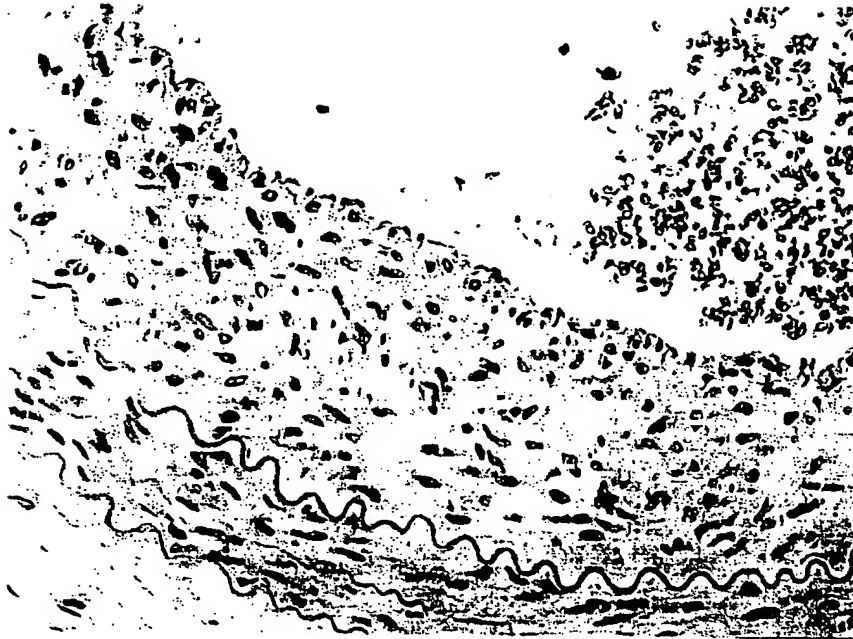
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FIGURE 9

Ep1.B enhances mitogenic stimulation



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FIGURE 10

A



B

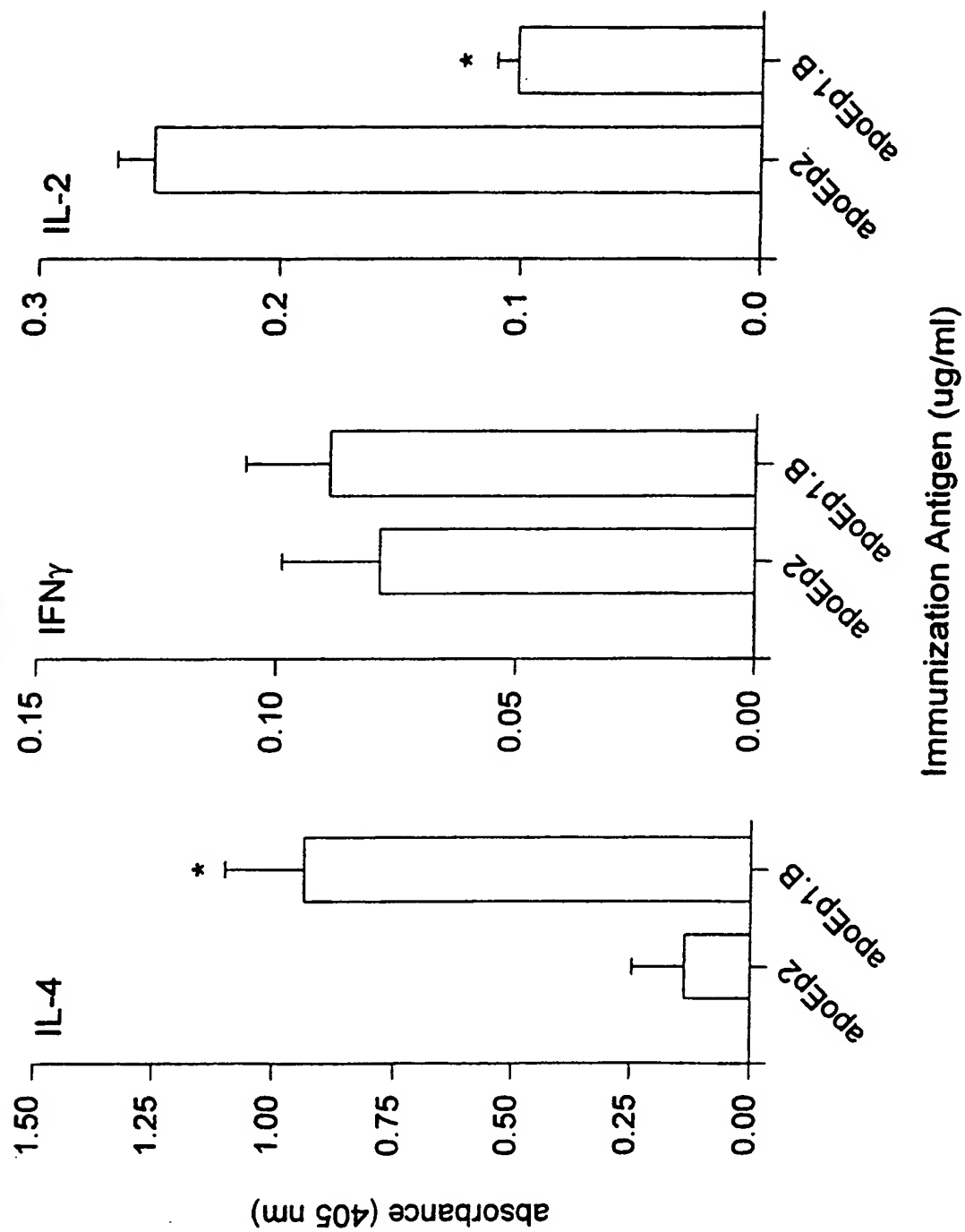


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FIGURE 10 cont'd

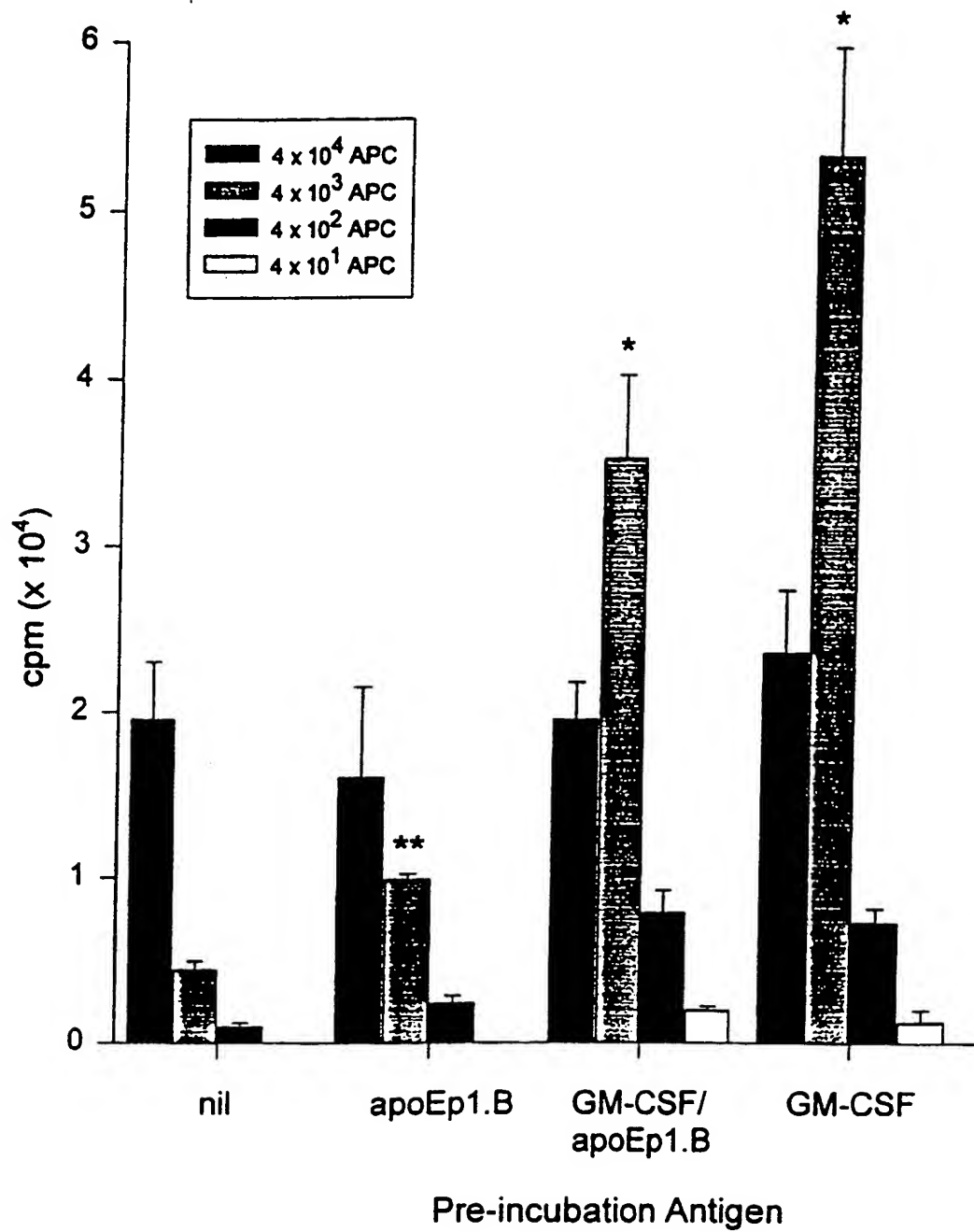
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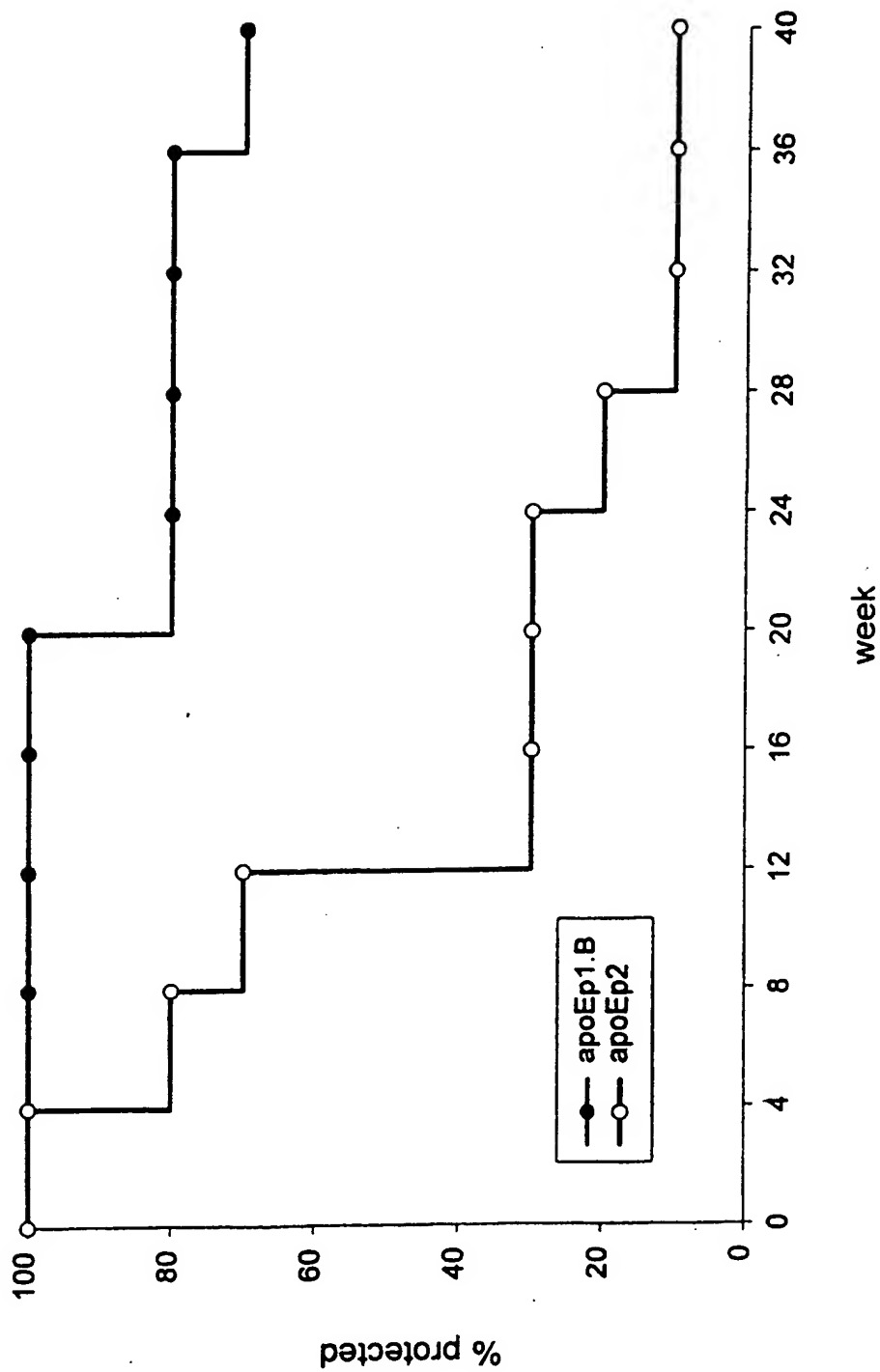


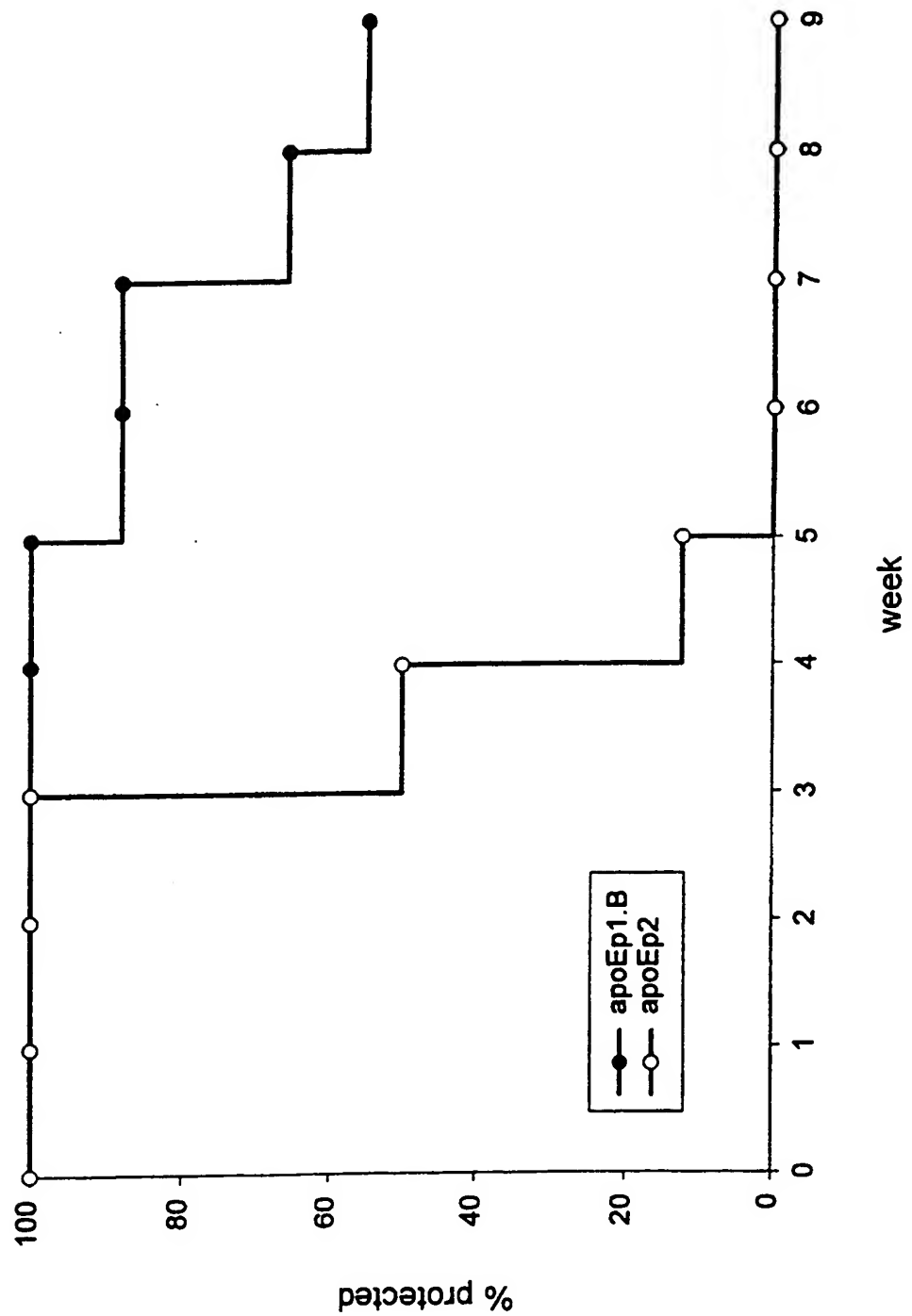
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FIGURE 11

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FIGURE 12

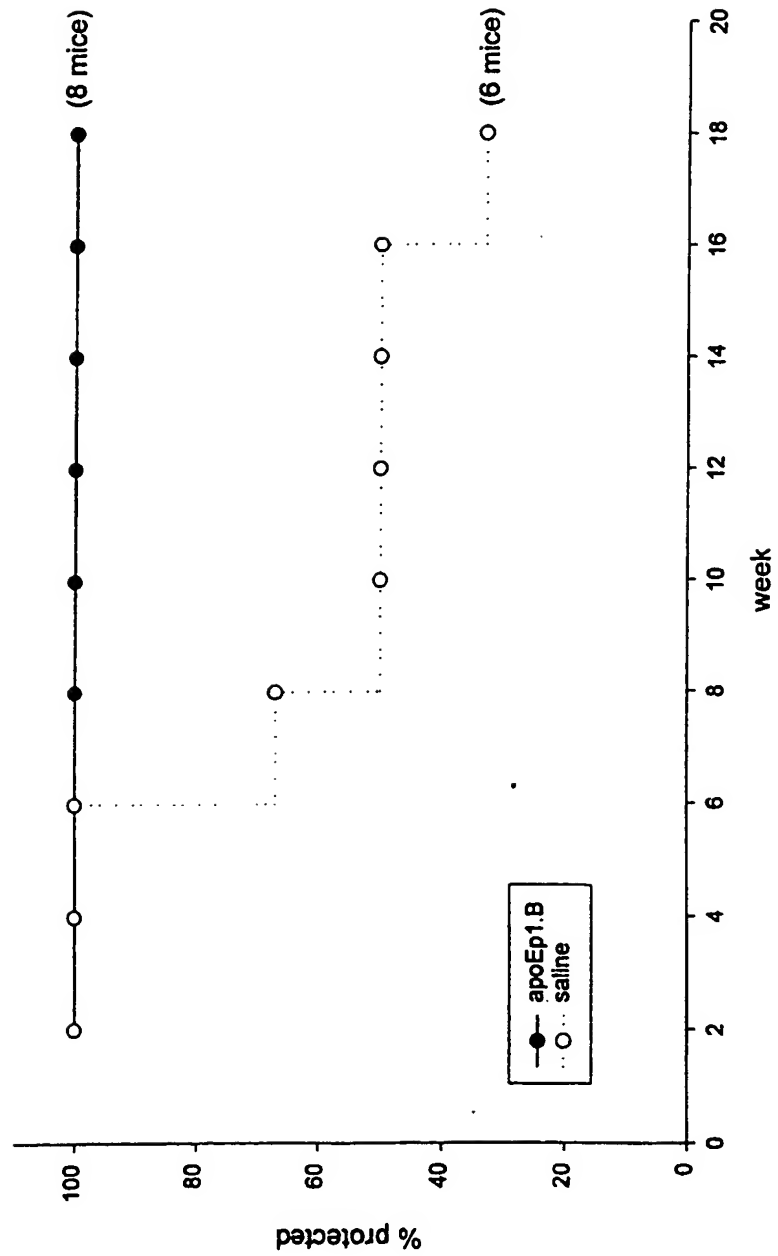


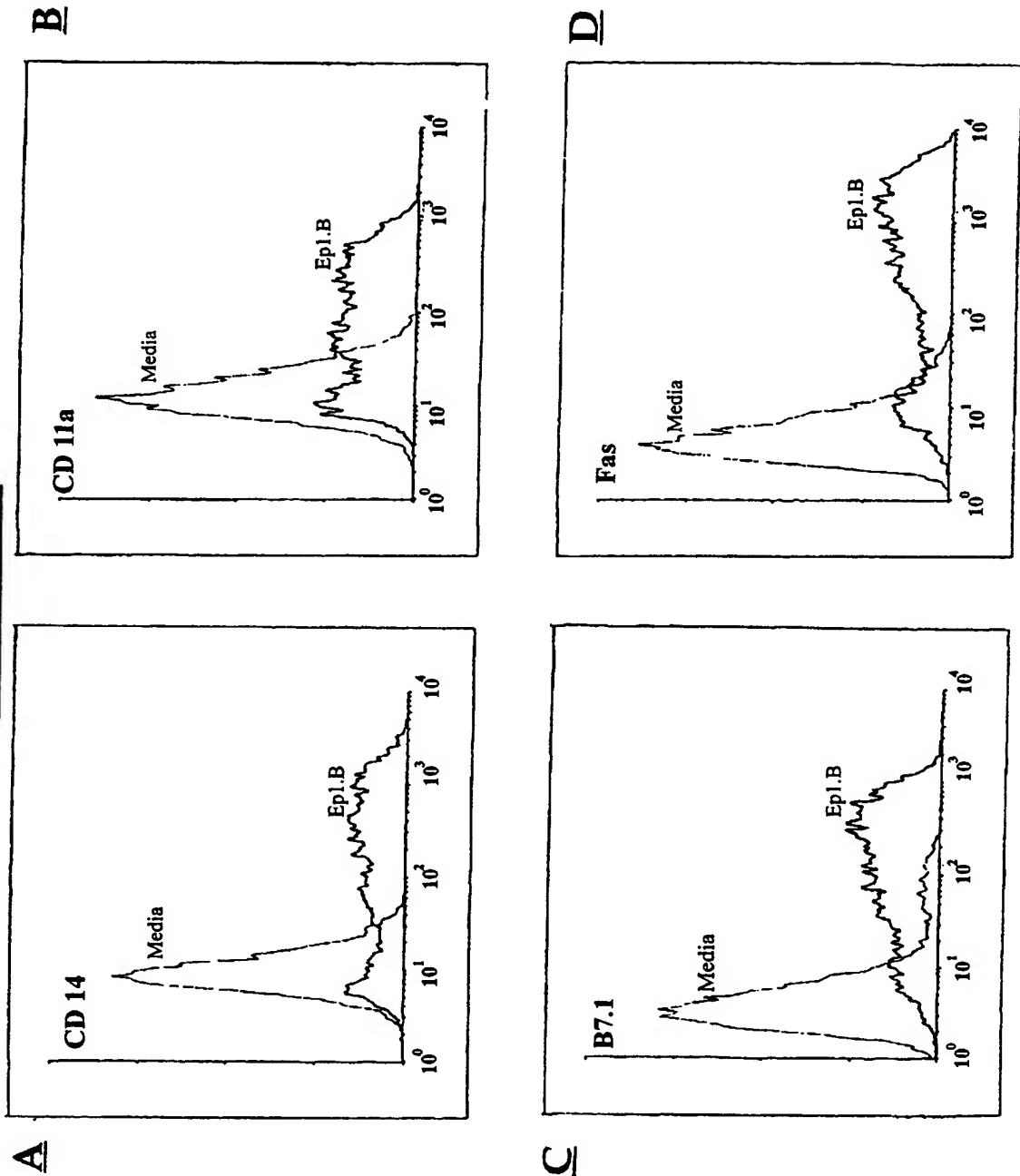
16/20FIGURE 13A

17/20FIGURE 13B

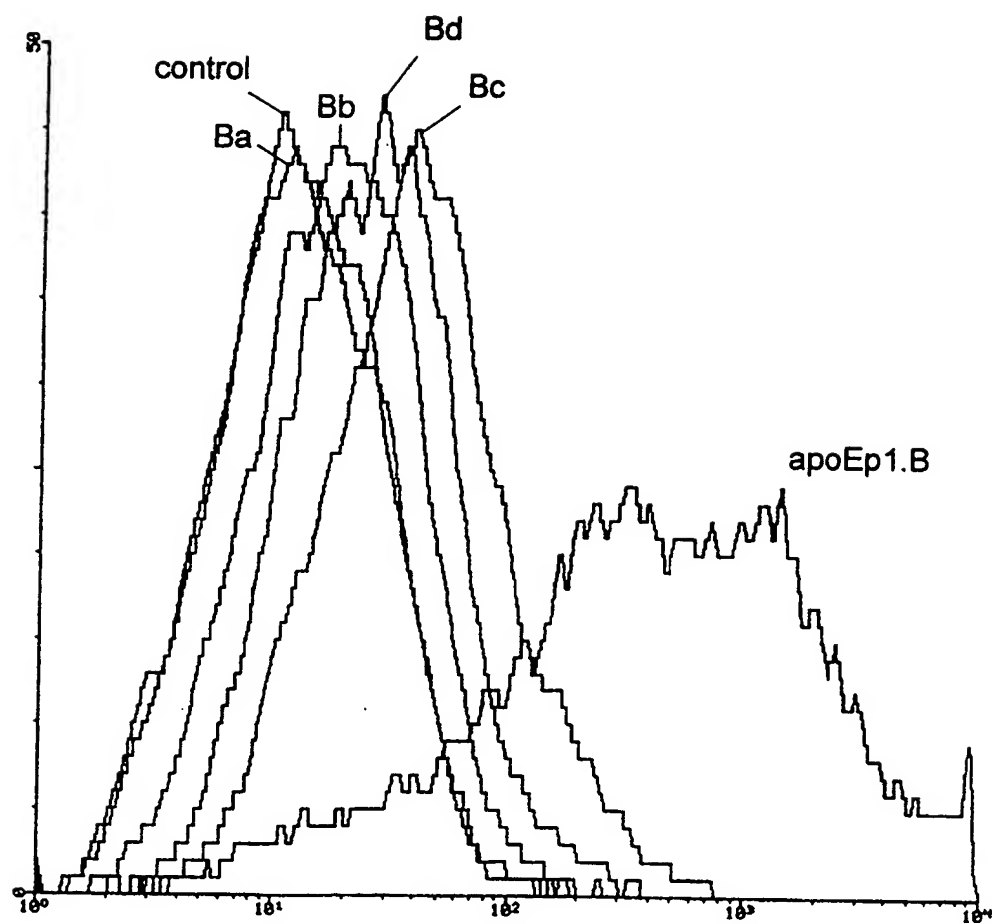
18/20FIGURE 14

ApoEp1.B footpad immunization in IFA protects NOD mice from diabetes



19/20FIGURE 15

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FIGURE 16



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SEQUENCE LISTING

<110> Rider, Beverley J.
Singh, Bhagirath

<120> Methods of Immune Modulation

<130> 9611-007

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2/2

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/12, C07K 14/775, A61K 38/17, 48/00	A3	(11) International Publication Number: WO 99/31227 (43) International Publication Date: 24 June 1999 (24.06.99)
(21) International Application Number: PCT/CA98/01129 (22) International Filing Date: 11 December 1998 (11.12.98) (30) Priority Data: 60/069,531 12 December 1997 (12.12.97) US (71) Applicant (for all designated States except US): THE UNIVERSITY OF WESTERN ONTARIO [CA/CA]; Stevenson - Lawson Building, Room 319, London, Ontario N6A 5B8 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): SINGH, Bhagirath [CA/CA]; 146 St. Bees Close, London, Ontario N6G 4B8 (CA). RIDER, Beverley [CA/US]; 531 Lake Boulevard, Redwood City, CA 94062 (US). (74) Agent: BERESKIN & PARR; 40th floor, 40 King Street West, Toronto, Ontario M5H 3Y2 (CA).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> (88) Date of publication of the international search report: 29 July 1999 (29.07.99)
(54) Title: NOVEL PEPTIDE, apoEp1.B, COMPOSITIONS AND USES THEREOF (57) Abstract <p>A peptide derived from apolipoprotein E termed apoEp1.B which includes amino acids 239-252 of the apolipoprotein E is described. The apoEp1.B peptide is a potent immune modulator that acts on a variety of immune cells. Interestingly, apoEp1.B is a dual modulator, capable of both inducing and suppressing an immune response. In particular, apoEp1.B has been shown to induce differentiation of stem cells into dendritic cells, to induce tumor cell differentiation and activation, to inhibit inflammation and to inhibit autoimmune disease.</p>		

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IPC 6 C12N15/12 C07K14/775 A61K38/17 A61K48/00

IPC 6 C07K

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	HUNT, DONALD F. ET AL: "Peptides presented to the immune system by the murine class II major histocompatibility complex molecule I-Ad"	33,34
A	SCIENCE (WASHINGTON, D. C., 1883-) (1992), 256(5065), 1817-20 CODEN: SCIEAS;ISSN: 0036-8075,1992, XP002104938 see page 1817 see abstract see page 1819; tables 1,2 see page 1820, paragraph 2 - paragraph 3 --- -/--	

☐ Patent family members are listed in annex.

"&" document member of the same patent family

17/06/1999

Sitch, W

INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 98/01129

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>DATABASE CHEMICAL ABSTRACTS ABSTRACT NO. 125:193352, RIDER, BEVERLEY J. ET AL: "Immune responses to self peptides naturally presented by murine class II major histocompatibility complex molecules" XP002104939 see abstract & MOL. IMMUNOL., vol. 33, no. 7/8, 1996, pages 625-633, -----</p>	

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA 98/01129

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 1-11 and 14-22 (all partially) and claim 13 (completely) are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
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Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

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